



The role of nitrogen as a driver of harmful algal blooms

Silvia Newell

Co-authors: Justyna Hampel, Daniel Hoffman, Mark McCarthy, Justin Myers, Tim Davis, Tom Johengen, Duane Gossieux



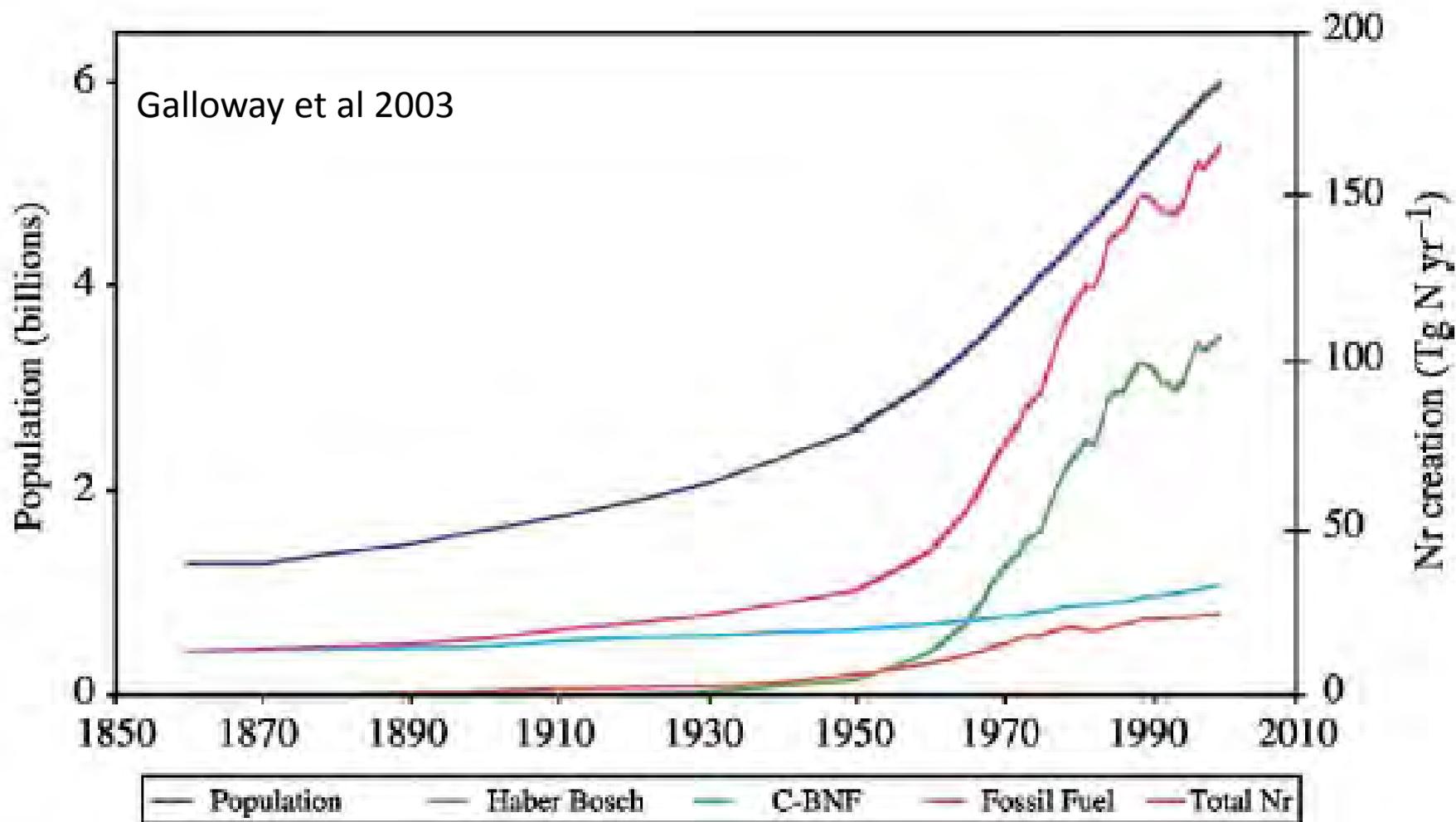
@NewellLab

Ecosystem impacts from Nutrient loading

- Harmful Algal Blooms
- Toxin production
- Fish Kills
- Oxygen depletion
- Greenhouse gas production



Anthropogenic N \geq Biological N fixation



Oct. 2008

Control
(no nutrients)



+ N-NO₃⁻



+ P-PO₄³⁻

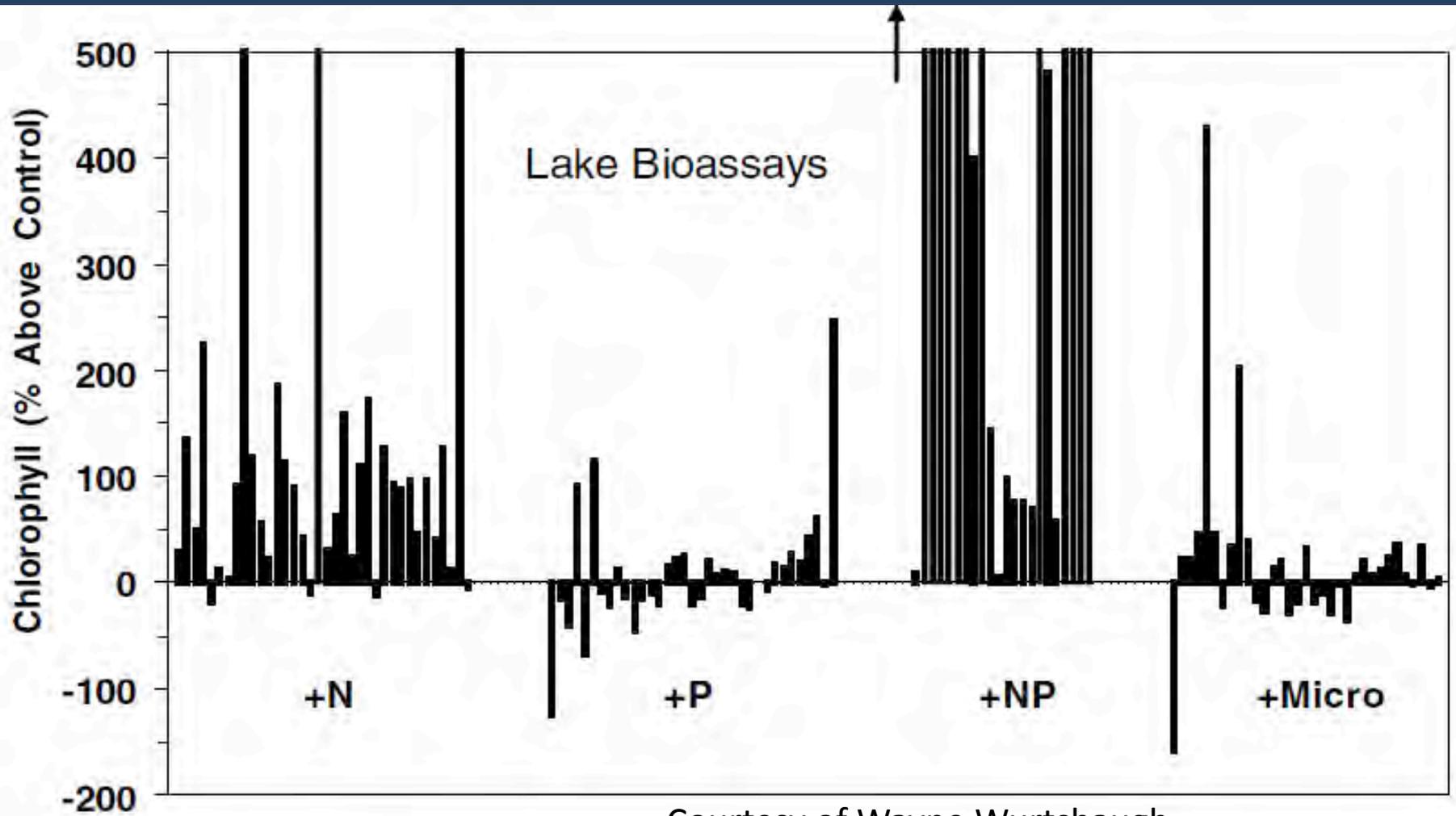


+ N + P

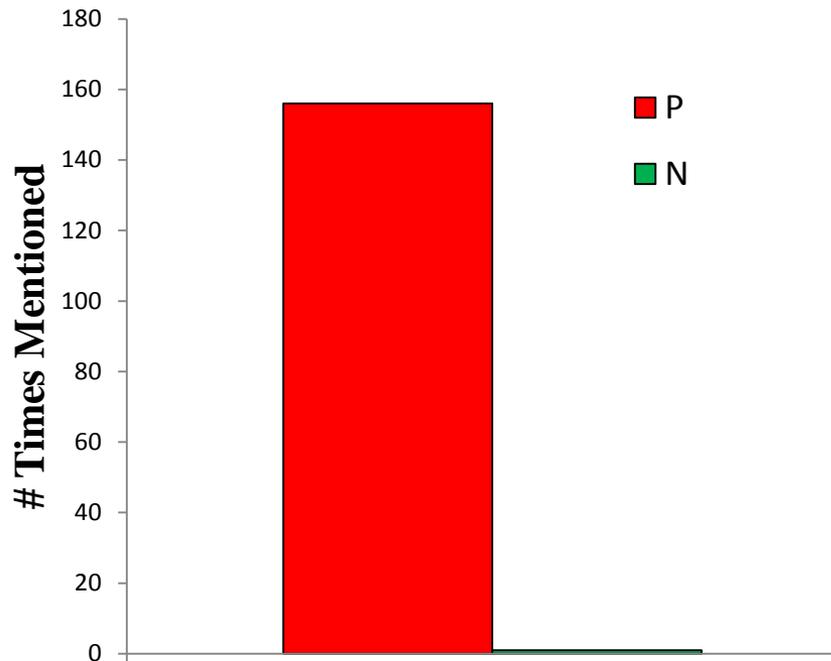


Courtesy of Hans Paerl

Nutrient Addition experiments



Courtesy of Wayne Wurtsbaugh



Almost 6 per page of text

Cyanobacteria = 16

Microcystis = 5

ARTICLE IN PRESS
JGLR-00684; No. of pages: 21; 4C: 3, 7, 12, 13, 16
Journal of Great Lakes Research xxx (2014) xxx-xxxx



ELSEVIER

Contents lists available at ScienceDirect

Journal of Great Lakes Research

journal homepage: www.elsevier.com/locate/jglr



Review

Assessing and addressing the re-eutrophication of Lake Erie: Central basin hypoxia

Donald Scavia^{a,*}, J. David Allan^b, Kristin K. Arend^c, Steven Bartell^d, Dmitry Beletsky^e, Nate S. Bosch^f, Stephen B. Brandt^g, Ruth D. Briland^h, Irem Dalogluⁱ, Joseph V. DePinto^j, David M. Dolan^k, Mary Anne Evans^k, Troy M. Farmer^h, Daisuke Goto^l, Haejin Han^m, Tomas O. Höökⁿ, Roger Knight^o, Stuart A. Ludsin^h, Doran Mason^p, Anna M. Michalak^q, R. Peter Richards^r, James J. Roberts^s, Daniel K. Rucinski^{h,t}, Edward Rutherford^p, David J. Schwab^l, Timothy M. Sesterhenn^u, Hongyan Zhang^e, Yuntao Zhou^{q,u}

^a Graham Sustainability Institute, University of Michigan, 625 E. Liberty, Ann Arbor, MI 48103, USA
^b School of Natural Resources and Environment, University of Michigan, 440 Church St., Ann Arbor, MI 48109, USA
^c Old Woman Creek National Estuarine Research Reserve, Ohio Department of Natural Resources, Division of Wildlife, Huron, OH 44839, USA
^d Cardno EM&E, 339 Whitecrest Dr., Maryville, TN 37801, USA
^e Cooperative Institute for Limnology and Ecosystems Research, School of Natural Resources and Environment, University of Michigan, 440 Church St., Ann Arbor, MI 48109, USA
^f Environmental Science, Grace College, Winona Lake, IN 46590, USA
^g Department of Fisheries and Wildlife, Oregon State University, Corvallis, OR 97331, USA
^h Aquatic Ecology Laboratory, Department of Evolution, Ecology, and Organismal Biology, The Ohio State University, 1314 Kinnear Rd., Columbus, OH 43212, USA
ⁱ Limnolab, 301 Aviz Drive, Ann Arbor, MI 484108, USA
^j University of Wisconsin-Green Bay, 2420 Nicolet Dr., Green Bay, WI, USA
^k U.S. Geological Survey, Great Lakes Science Center, 1451 Green Rd., Ann Arbor, MI 48105, USA
^l Center for Limnology, University of Wisconsin-Madison, 680 North Park Street, Madison, WI 53706, USA
^m Korea Environment Institute, 215 Jirheung-ro, Eunpyeong-gu, Seoul 122-706, Republic of Korea
ⁿ Department of Forestry and Natural Resources, Purdue University, 196 Mandlel St. West Lafayette, IN 47907, USA
^o Division of Wildlife, Ohio Department of Natural Resources, Columbus, OH 43229, USA
^p Great Lakes Environmental Research Laboratory, NOAA, 4940 S. State Rd., Ann Arbor, MI 48108, USA
^q Department of Global Ecology, Carnegie Institute for Science, 260 Panama St., Stanford, CA 94305, USA
^r National Center for Water Quality Research, Heidelberg University, 310 E. Market St., Tiffin, OH 44883, USA
^s U.S. Geological Survey, Fort Collins Science Center, 2150 Centre Ave., Fort Collins, CO 80523, USA
^t Water Center, University of Michigan, 625 E. Liberty, Ann Arbor, MI 48103, USA
^u Department of Civil and Environmental Engineering, University of Michigan, Ann Arbor, MI 48109, USA

ARTICLE INFO

Article history:
 Received 14 September 2013
 Accepted 17 January 2014
 Available online xxxxx

Communicated by Leon Boegman

Keywords:
 Lake Erie
 Hypoxia
 Phosphorus load targets
 Best management practices

ABSTRACT

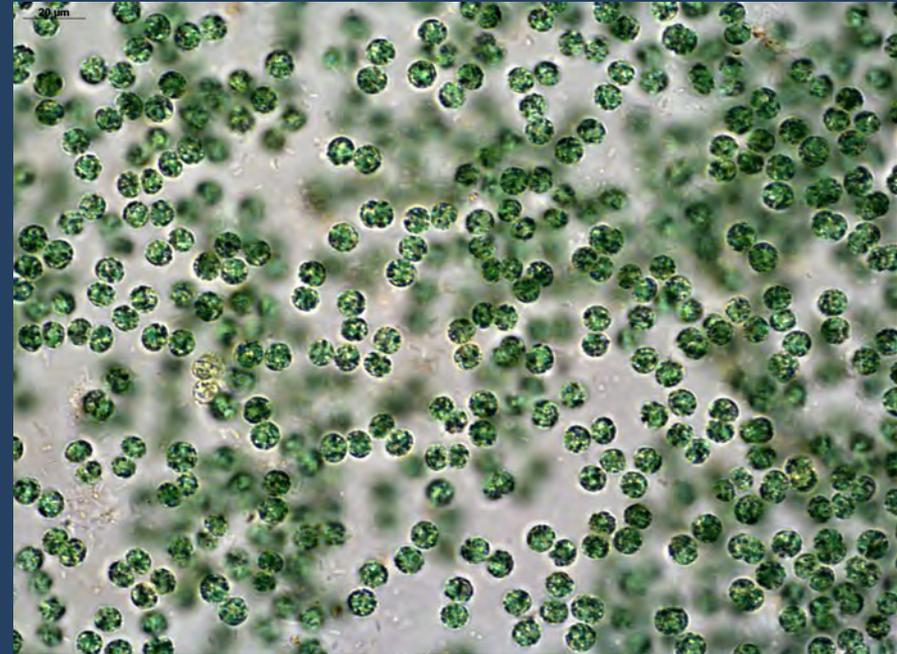
Relieving phosphorus loading is a key management tool for controlling Lake Erie eutrophication. During the 1960s and 1970s, increased phosphorus inputs degraded water quality and reduced central basin hypolimnetic oxygen levels which, in turn, eliminated thermal habitat vital to cold-water organisms and contributed to the extirpation of important benthic macroinvertebrate prey species for fishes. In response to load reductions initiated in 1972, Lake Erie responded quickly with reduced water-column phosphorus concentrations, phytoplankton biomass, and bottom-water hypoxia (dissolved oxygen <2 mg/l). Since the mid-1990s, cyanobacteria blooms increased and extensive hypoxia and benthic algae returned. We synthesize recent research leading to guidance for addressing this re-eutrophication, with particular emphasis on central basin hypoxia. We document recent trends in key eutrophication-related properties, assess their likely ecological impacts, and develop load response curves to guide revised hypoxia-based loading targets called for in the 2012 Great Lakes Water Quality Agreement. Reducing central basin hypoxic area to levels observed in the early 1990s (ca. 2000 km²) requires cutting total phosphorus loads by 46% from the 2003–2011 average or reducing dissolved reactive phosphorus loads by 78% from the 2005–2011 average. Reductions to these levels are also protective of fish habitat. We provide potential approaches for achieving those new loading targets, and suggest that recent load reduction recommendations focused on western basin cyanobacteria blooms may not be sufficient to reduce central basin hypoxia to 2000 km².

© 2014 International Association for Great Lakes Research. Published by Elsevier B.V. All rights reserved.

Community Dominance: Lake Erie cyanobacterial Harmful Algal Blooms

1960s and 1970s

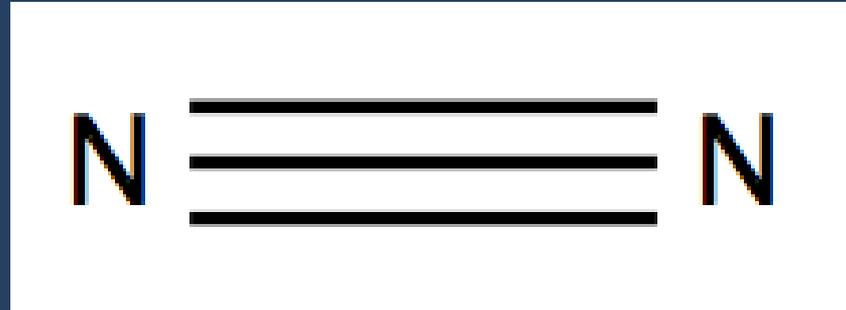
1990s to Present



© University of New
Hampshire

From
<http://cyanobacteria.myspecies.info>

N Bioavailability

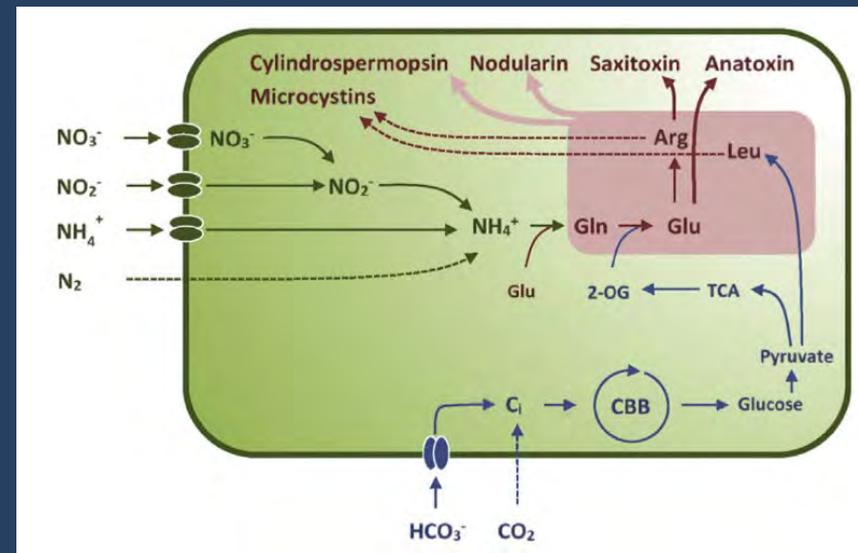
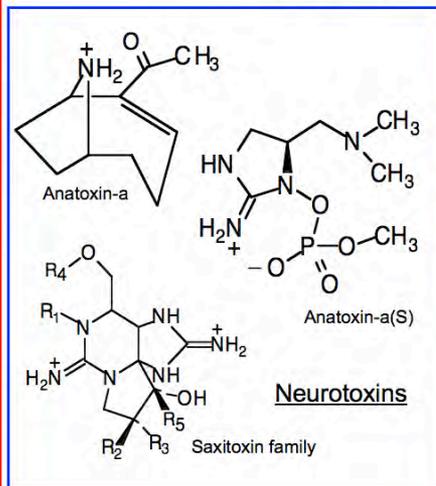
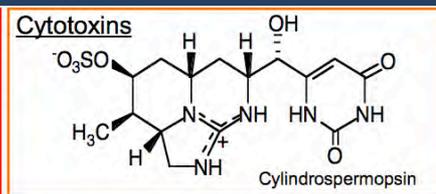
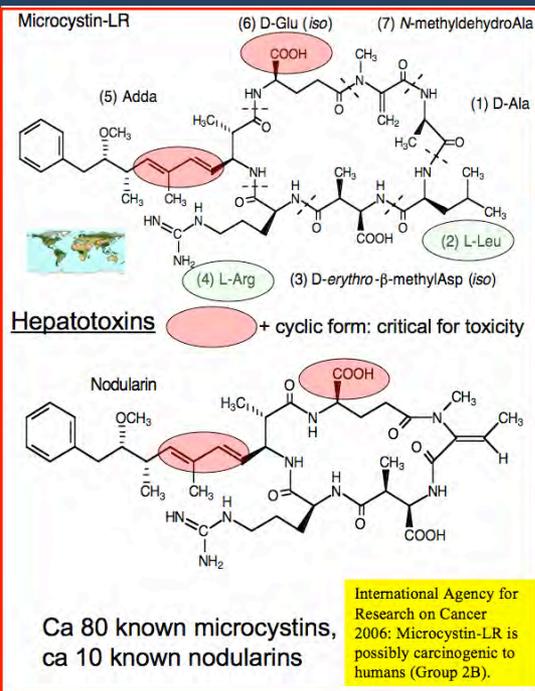


Triple bond -> difficult to break
18 ATP required for each N₂ molecule
N₂: limited bioavailability

More useful forms:



N and Cyanobacteria Toxicity



© Jussi Meriluoto

Gobler et al. 2016

N and Cyanobacteria Toxicity

- Reduced N form additions to non-N-fixing cyanobacteria can increase toxicity.

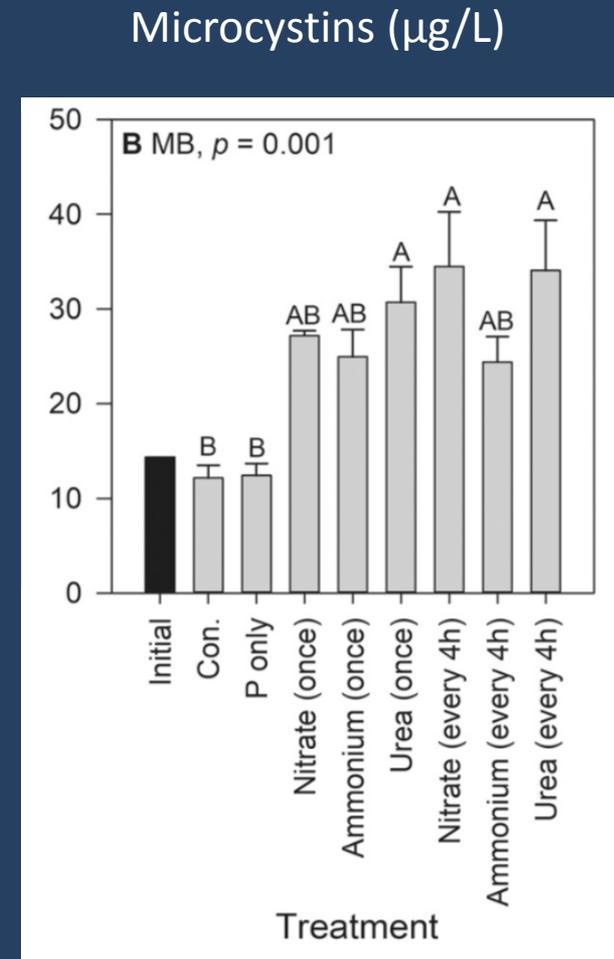
(Davis et al. 2010, 2015; Chaffin et al. 2018)

- Low NH_4^+ concentrations can inhibit toxin production

(Kuniyoshi et al. 2010)

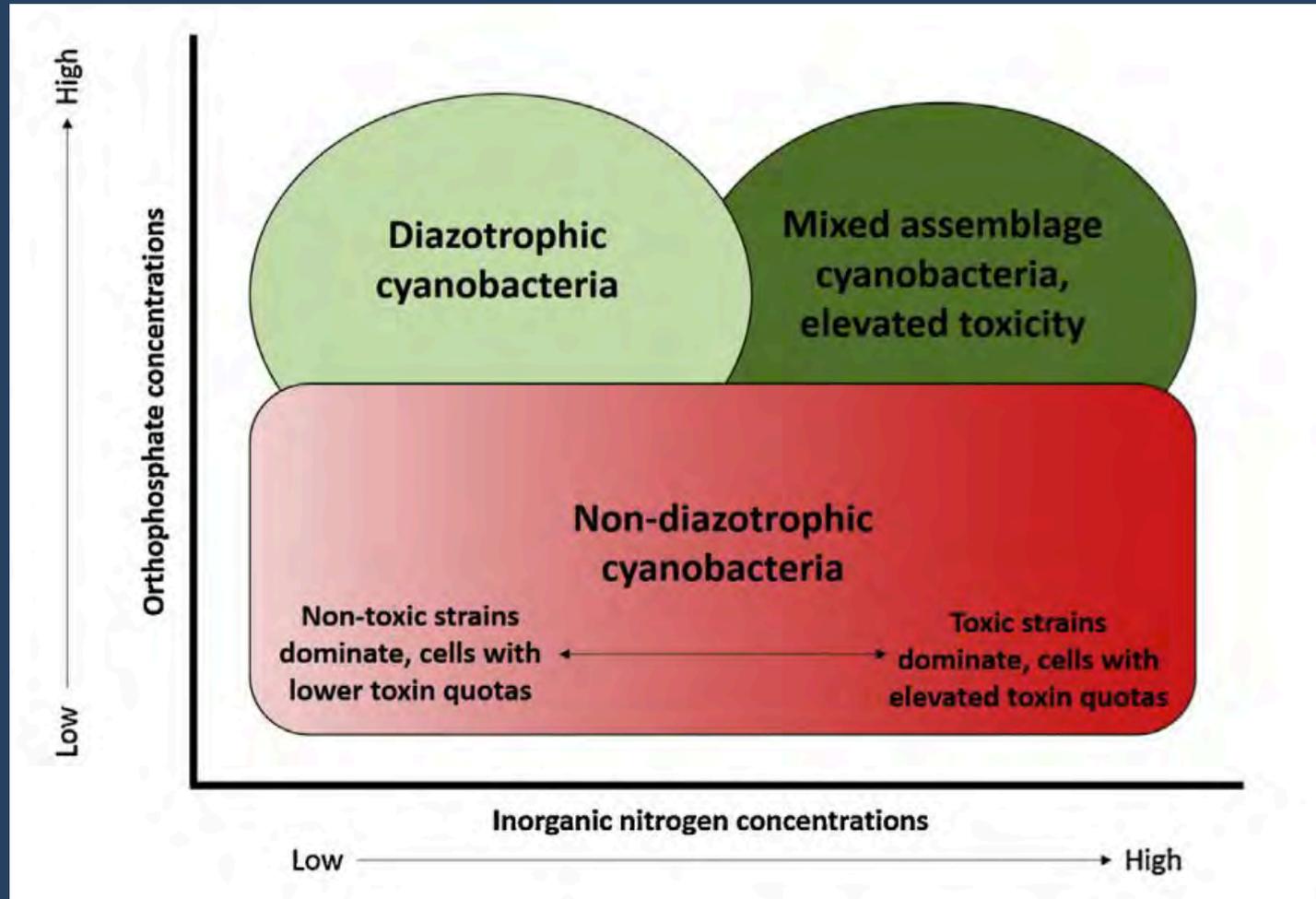
- NH_4^+ and urea uptake may lead to both increased *Microcystis* biomass and toxin production

(Harke et al. 2016)



Chaffin et al. 2018

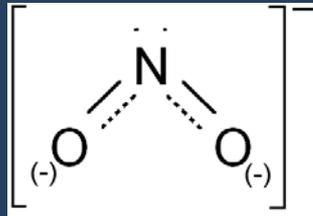
Implications for Community Shift



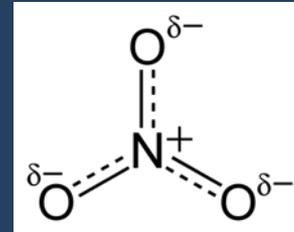
Nitrogen



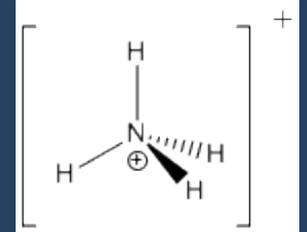
Dinitrogen Gas



Nitrite



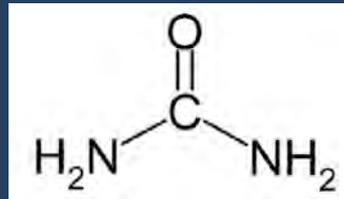
Nitrate



Ammonia/Ammonium

DON

Dissolved Organic N



Urea



Nitrous Oxide

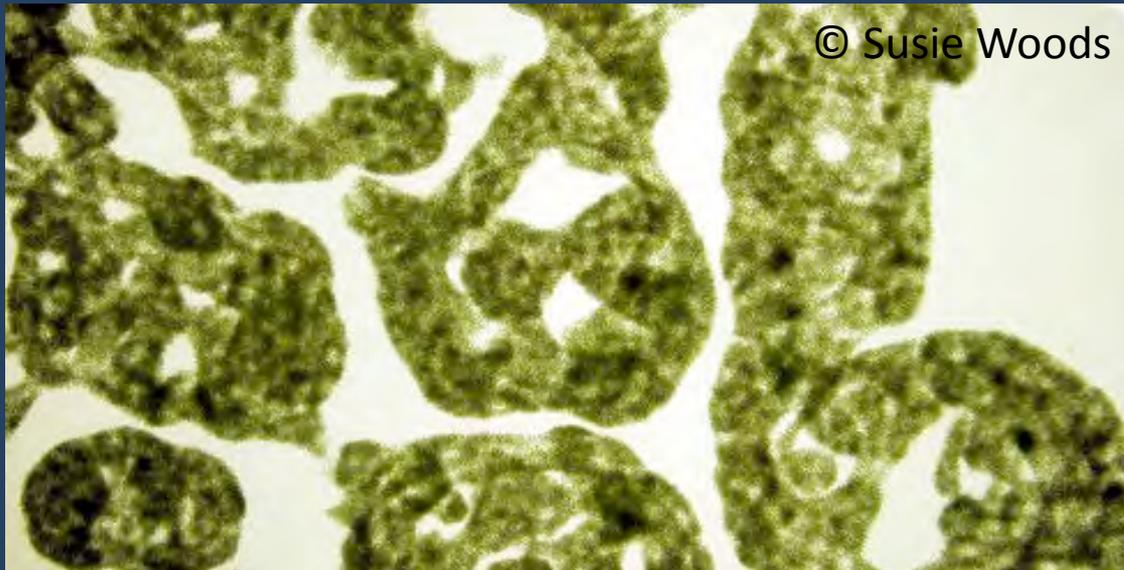
DIN

Dissolved Inorganic N

Ammonium: the common currency

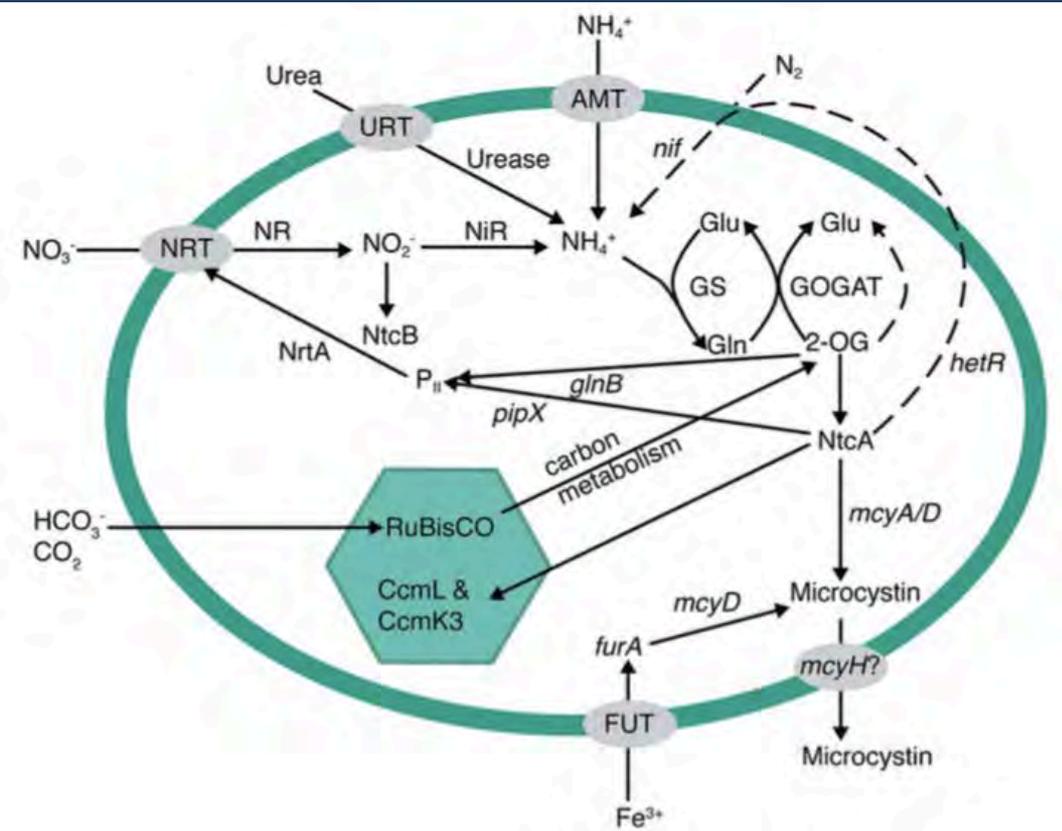


Diatoms use and store nitrate efficiently

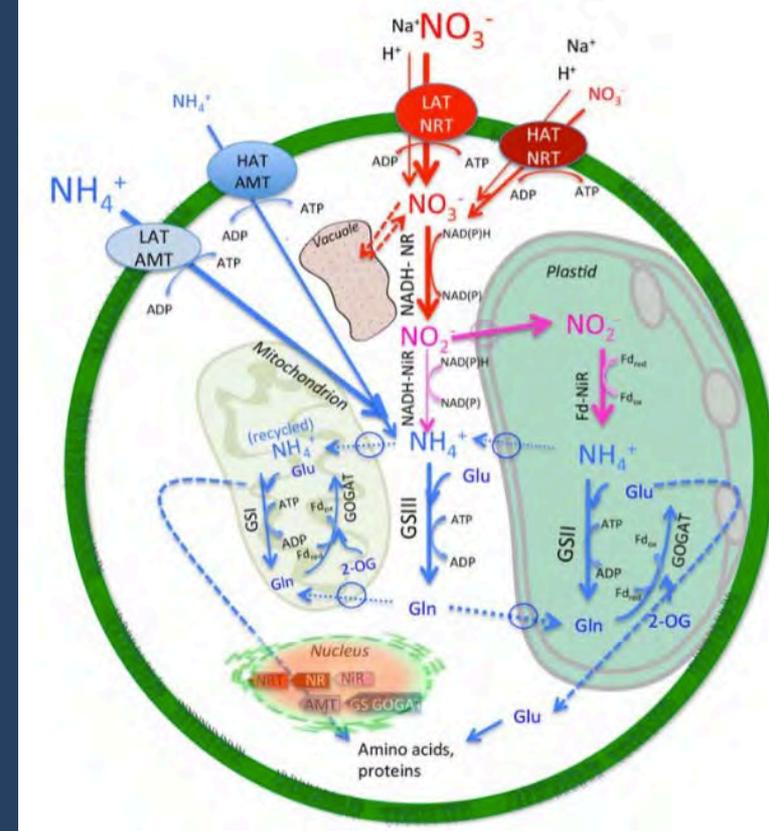


Most phyto
greatly prefer
& *Microcystis* has a
very strong affinity
for NH_4^+

N Assimilation in Phytoplankton



Beversdorf et al. 2015

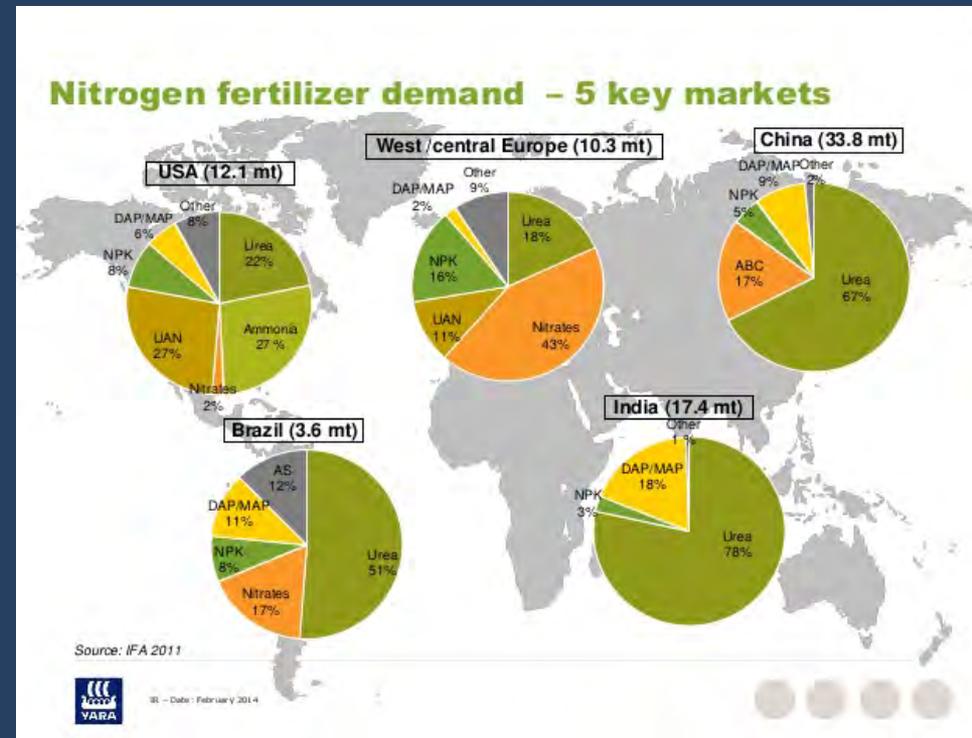


Glibert et al. 2015

N in Fertilizer

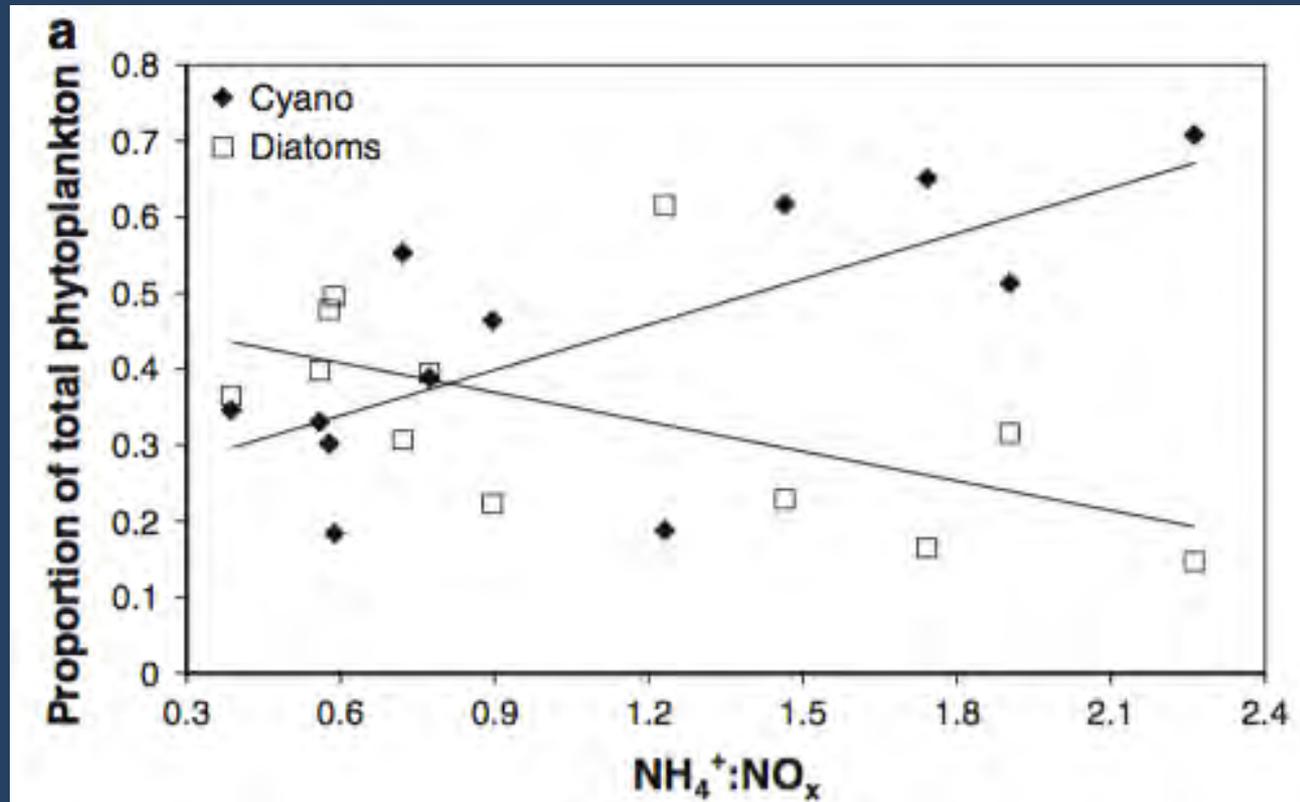
Global shift toward urea and/or anhydrous ammonia.

Urea = >50% of worldwide applications
(Glibert et al. 2006)



N Form and Community Structure

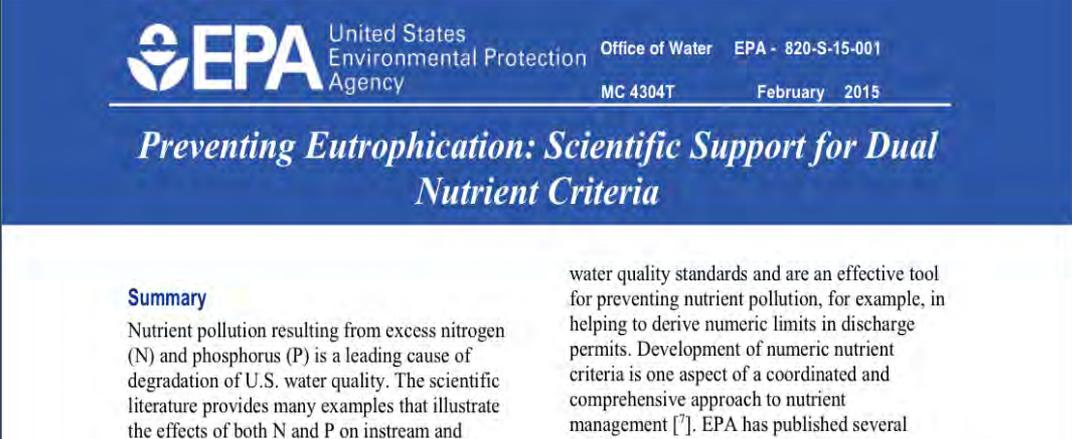
- NO_3^- : favors diatoms
- Reduced N (NH_4^+ and urea): favors cyanobacteria



(Limited) Regulations on N Inputs

- U.S. EPA Bulletin 820-S-15-001

Advocates for dual nutrient control strategy



The image shows the cover of an EPA bulletin. The top section is a dark blue header with the EPA logo on the left, the text 'United States Environmental Protection Agency' in the middle, and 'Office of Water EPA - 820-S-15-001' on the right. Below the header, the title 'Preventing Eutrophication: Scientific Support for Dual Nutrient Criteria' is written in a white, italicized serif font. The main body of the cover is white and contains a 'Summary' section. The summary text is partially visible on the left side of the image, and the right side of the summary text is cut off by the edge of the image.

Summary

Nutrient pollution resulting from excess nitrogen (N) and phosphorus (P) is a leading cause of degradation of U.S. water quality. The scientific literature provides many examples that illustrate the effects of both N and P on instream and

water quality standards and are an effective tool for preventing nutrient pollution, for example, in helping to derive numeric limits in discharge permits. Development of numeric nutrient criteria is one aspect of a coordinated and comprehensive approach to nutrient management [?]. EPA has published several

- No N reduction targets in Ohio

Proposed 40% reduction in P loading

Great Lakes Water Quality Agreement – Annex IV

Case study: Lake Erie



Maumee River

Increasing CyanoHABs in Western Lake Erie linked to increase in SRP

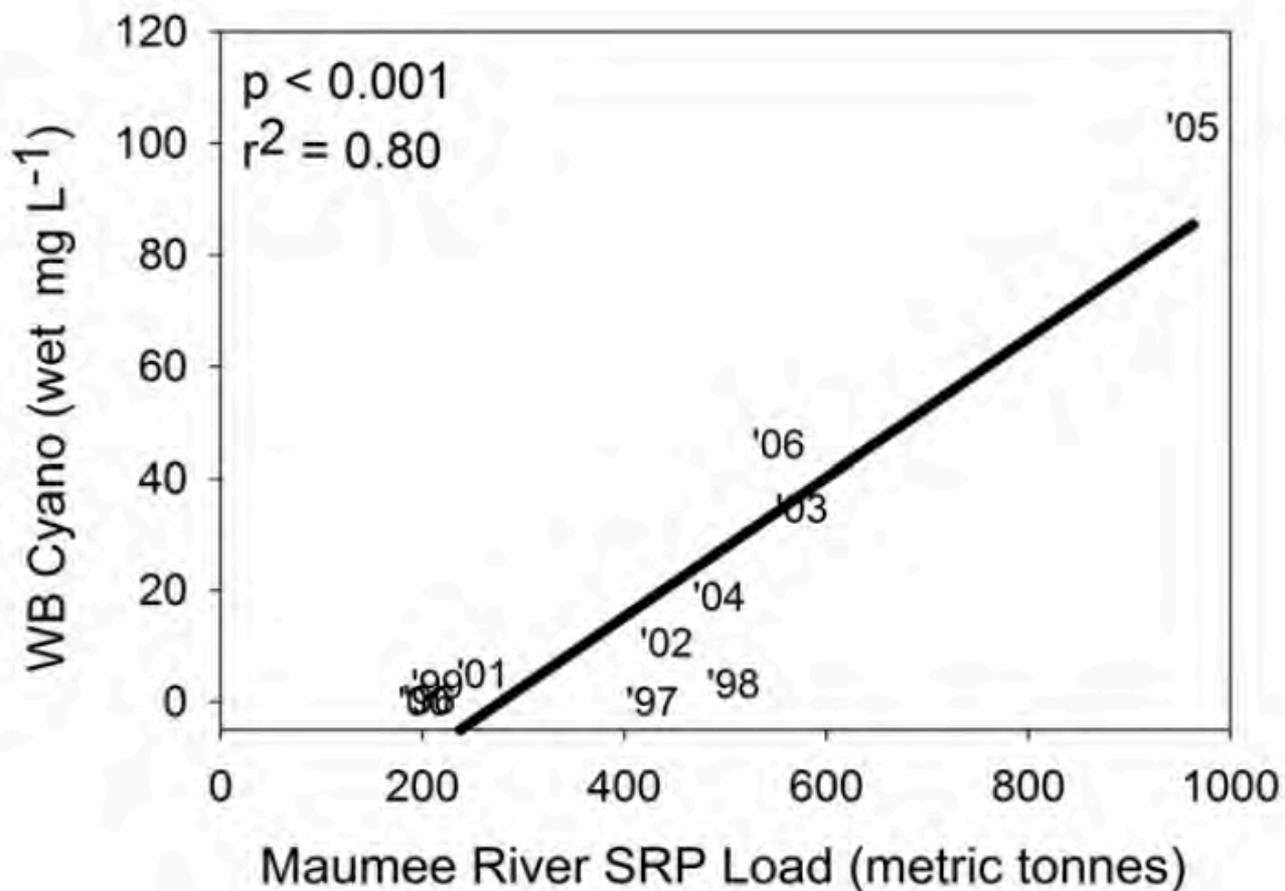
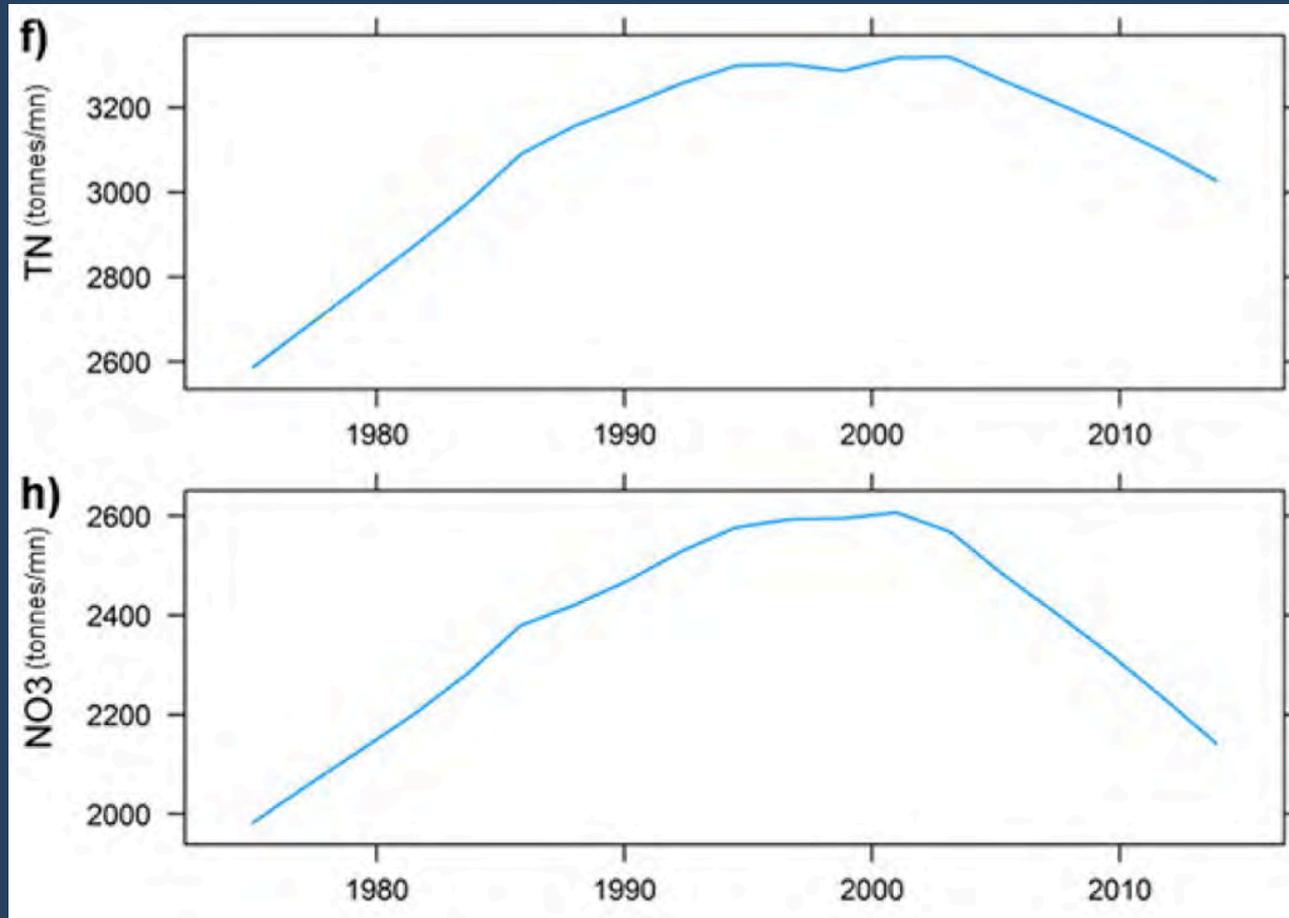


Fig. 4. Western basin (WB) median cyanobacteria wet-weight biomass (mg L^{-1}) (Cyano) as a function of Maumee River SRP load (metric tonnes) for water years 1996–2006.

Maumee N Loads to Western Lake Erie

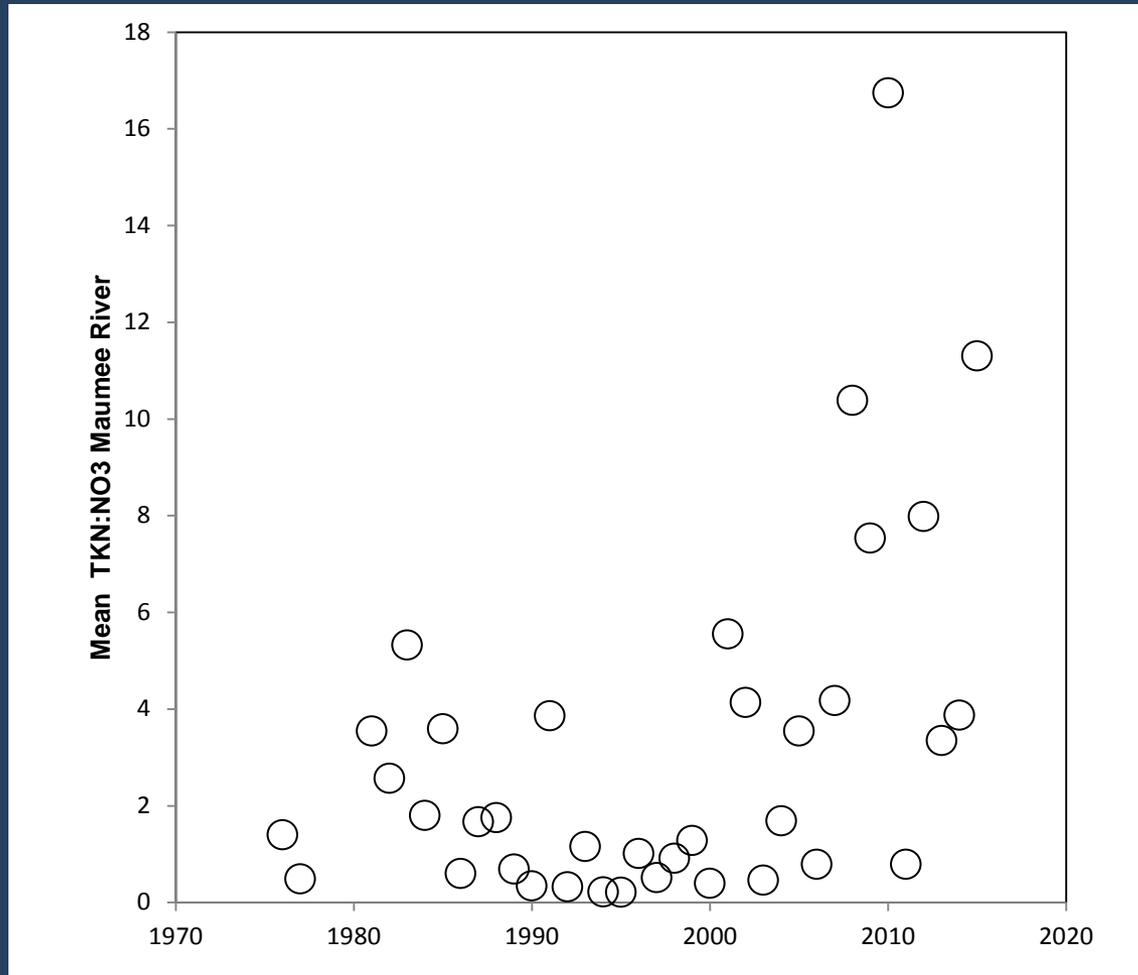


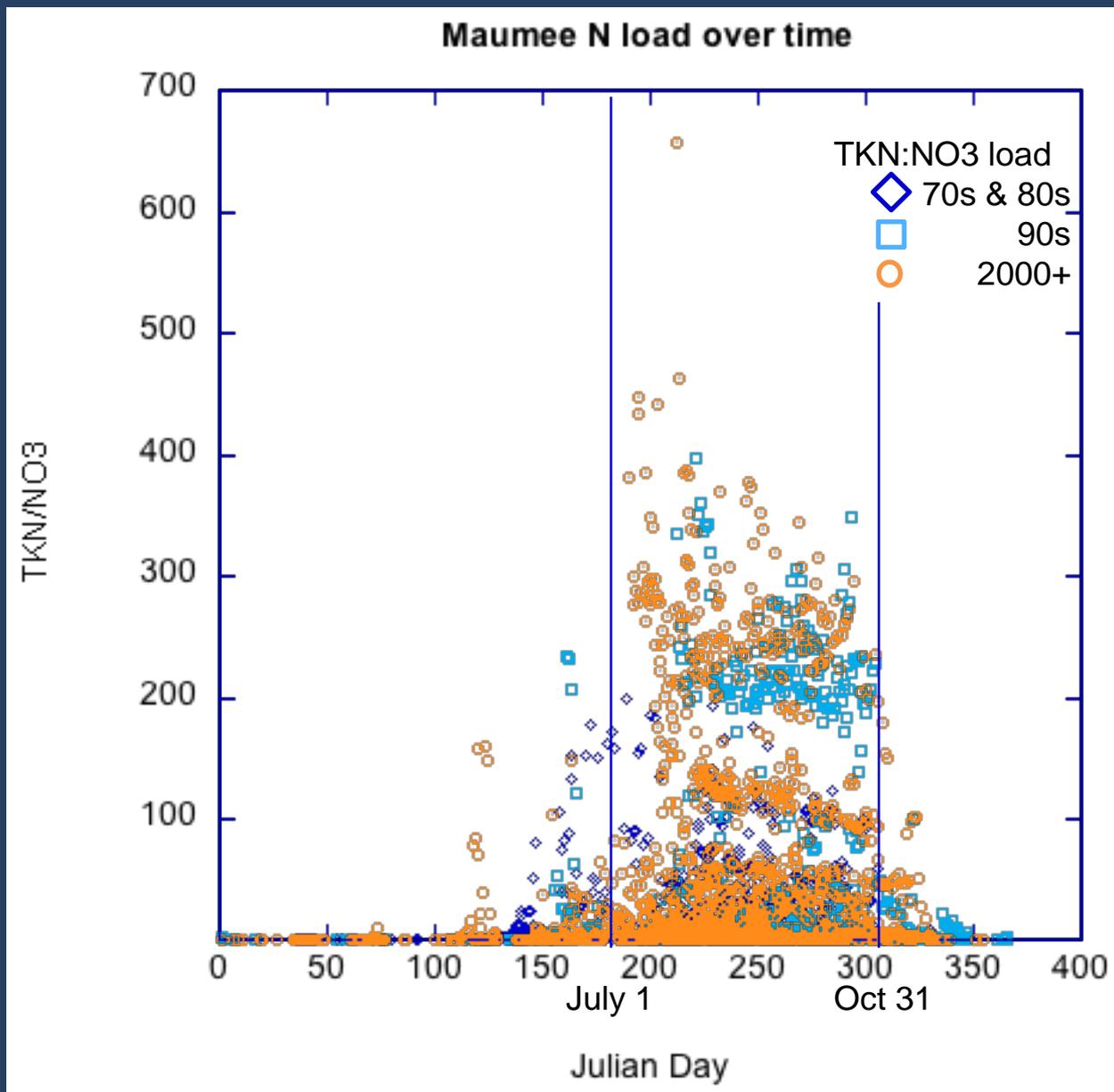
N Inputs to Western Lake Erie



- Maumee River: largest Great Lakes watershed
- Kjeldahl N (NH_4^+ + organic N) load from Maumee River to Lake Erie
= 9000 metric tons/yr
 $\frac{1}{4}$ of total N load
(Richards et al. 2010)

Reduced N % of Maumee Load increasing

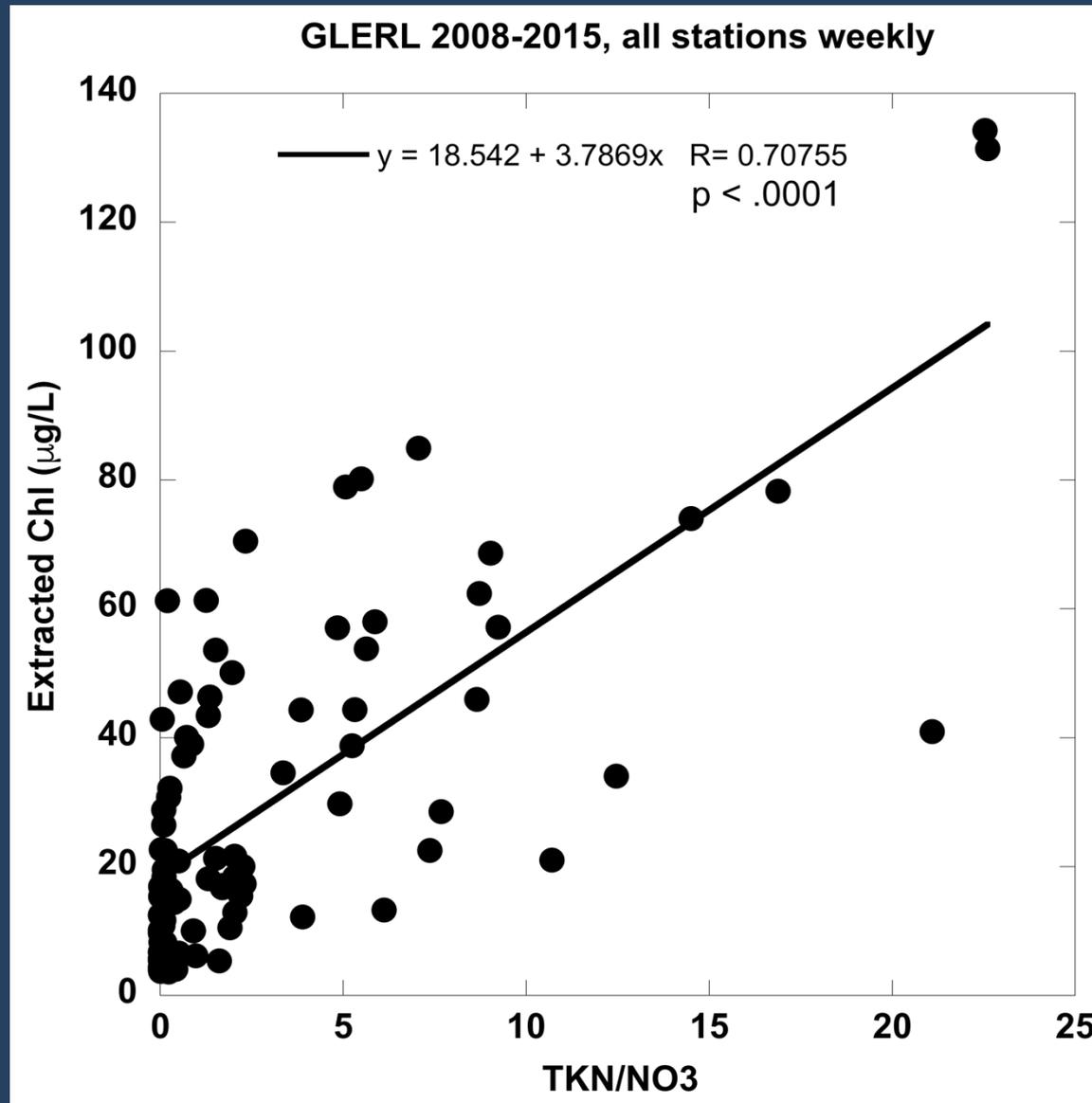




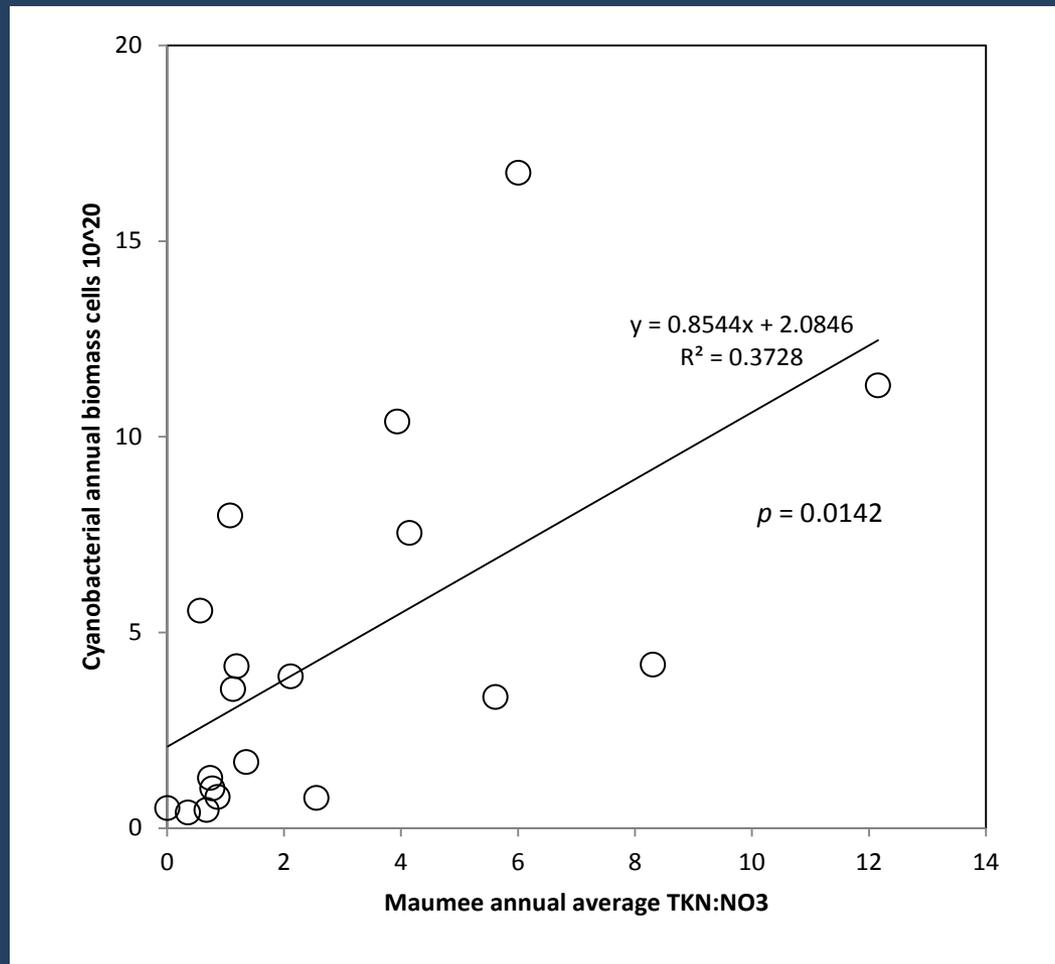
*Most values outside of summer are <1; y-axis scale starts at 1

Newell et al., 2019

TKN/NO₃⁻ vs Chlorophyll concentrations



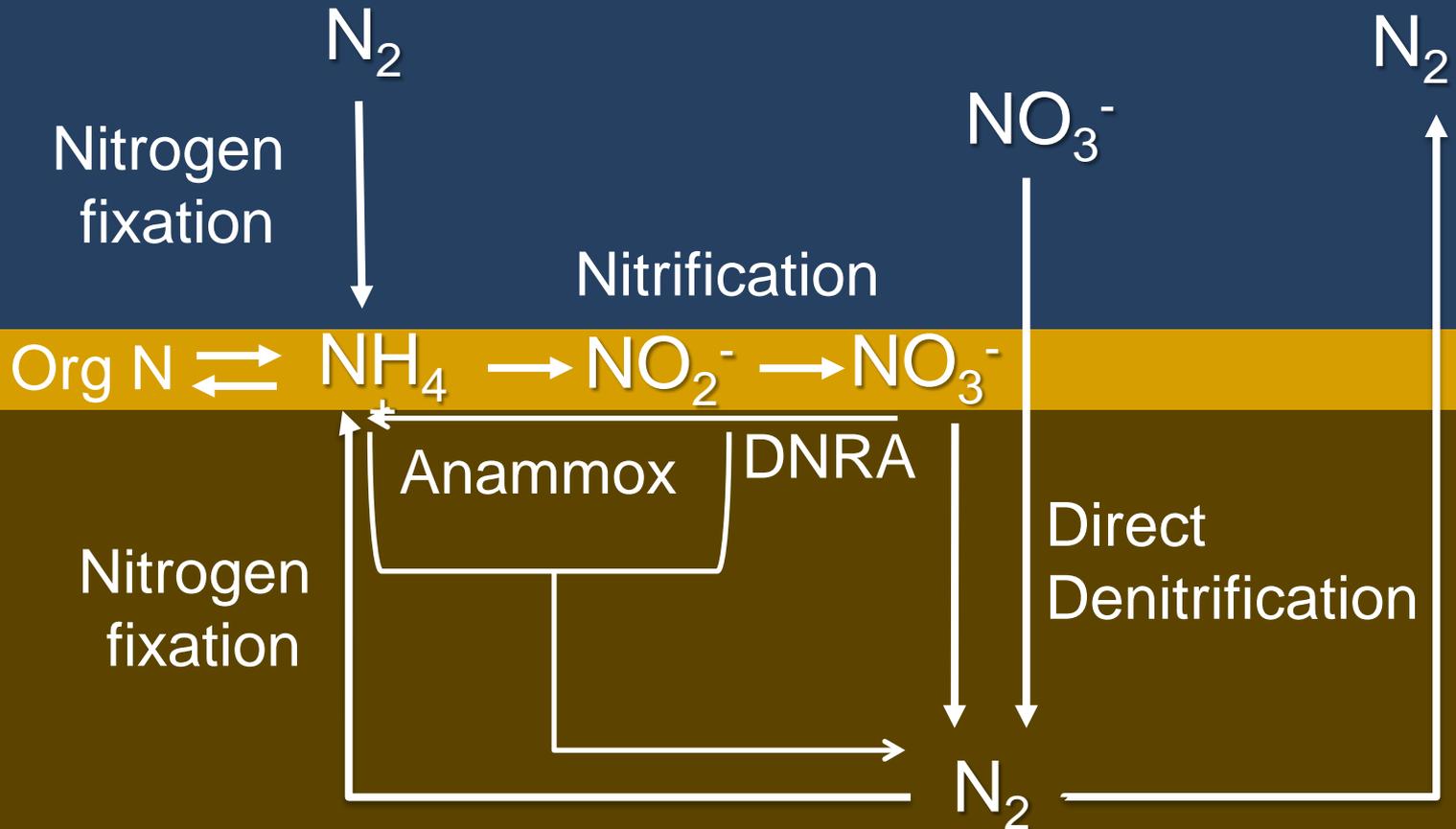
The proportion of reduced to oxidized N in the Maumee River load is significantly correlated to the increase in cyanobacterial bloom biomass in Western Lake Erie.





Nitrogen Cycle

water



sediment

Newell Lab research objectives

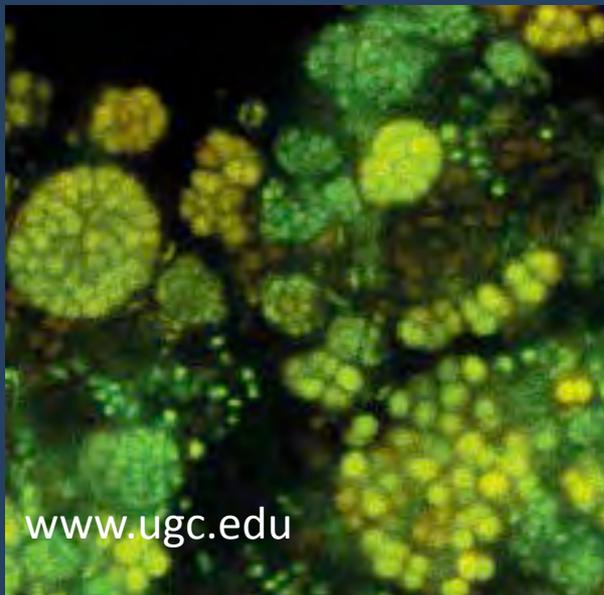
- To use a paired molecular-biogeochemical approach to disentangle the intricacies of the nitrogen cycle
- To understand the impact of human activities (primarily climate change and increased nitrogen loading) on the nitrogen cycle

Ammonium: the common currency



NH_4^+ half-saturation constant (K_m) for *Microcystis* is high:
0.5-37 μM

Nicklisch and Kohl 2007, Takeya et al. 2004



Ammonia-oxidizing bacteria can have a very high K_m (5-300 μM) but ammonia-oxidizing archaea have a very, very low K_m 0.05-0.15 μM

Internal N loading

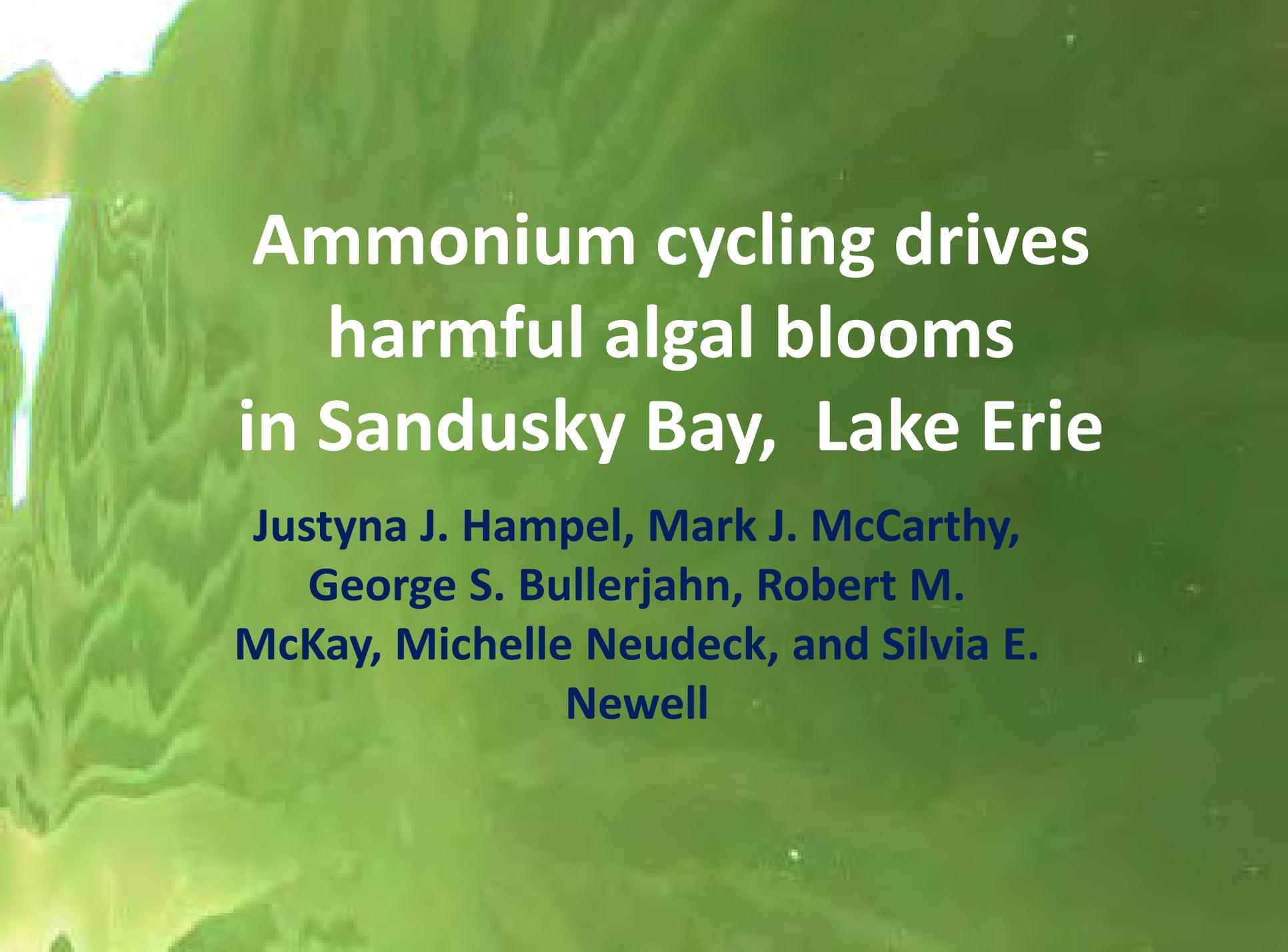
Research Questions

- How quickly is NH_4^+ recycled?
- To what extent can NH_4^+ recycling support cyanobacterial bloom growth?
- When do phytoplankton become N-limited?

N Inputs to Western Lake Erie



- Maumee River: largest Great Lakes watershed
- Kjeldahl N (NH_4^+ + organic N) load from Maumee River to Lake Erie
= 9000 metric tons/yr
 $\frac{1}{4}$ of total N load
(Richards et al. 2010)

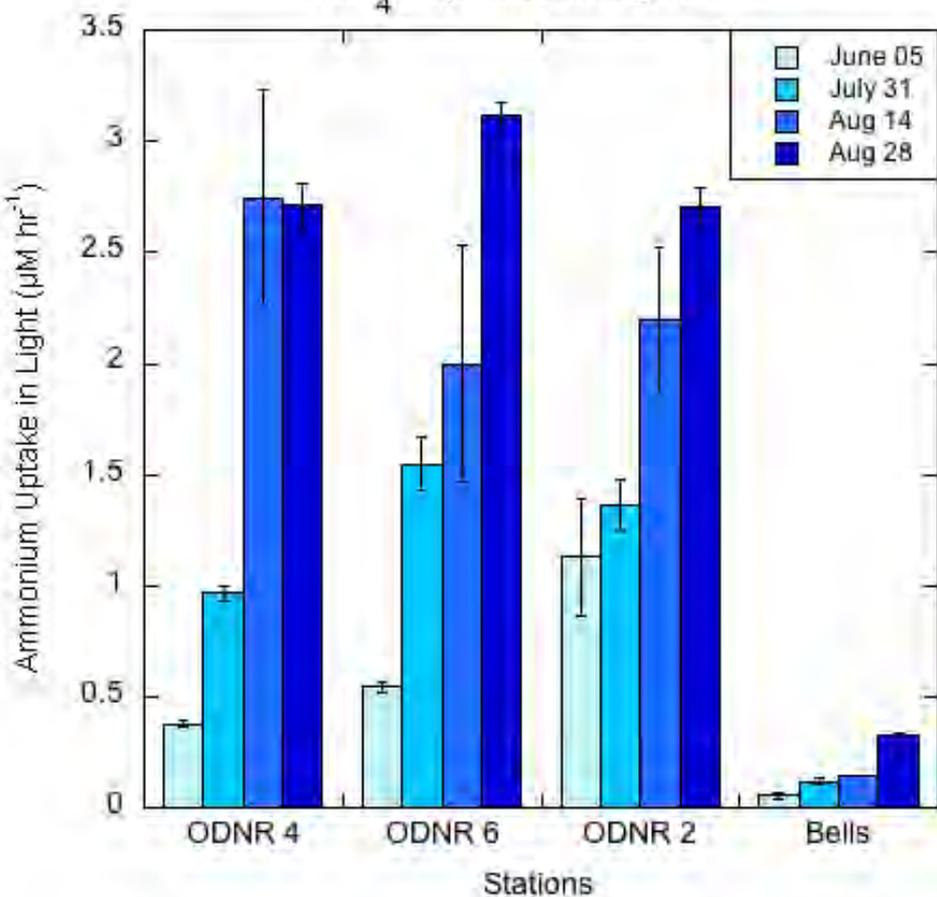


Ammonium cycling drives harmful algal blooms in Sandusky Bay, Lake Erie

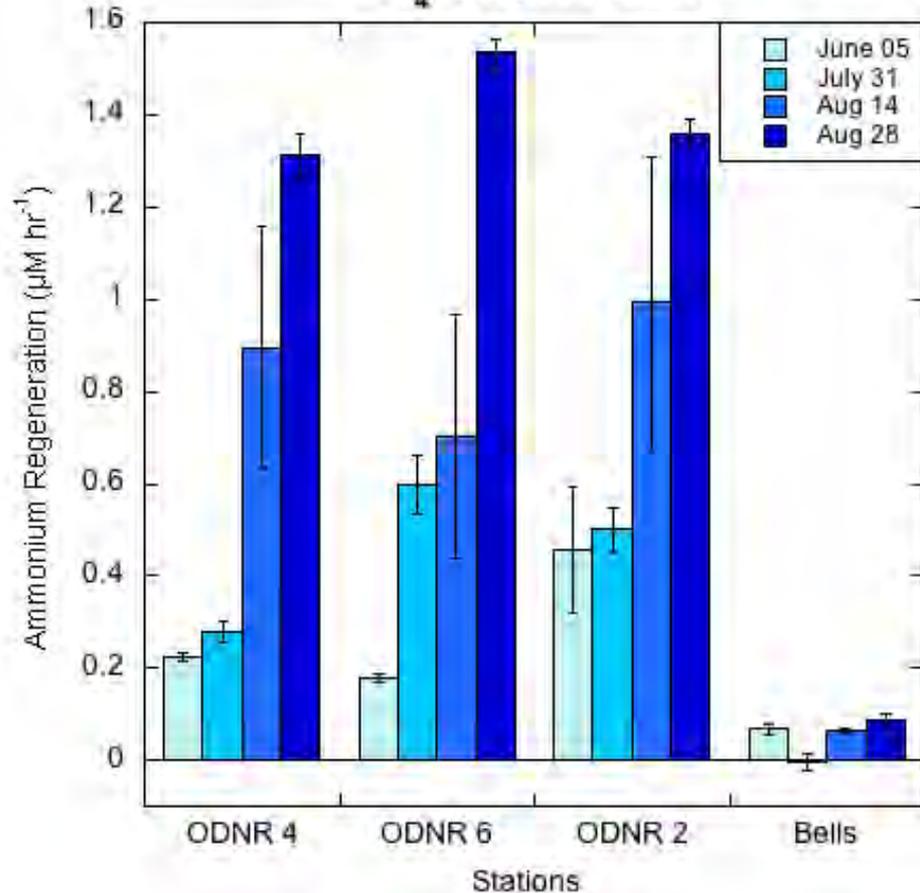
**Justyna J. Hampel, Mark J. McCarthy,
George S. Bullerjahn, Robert M.
McKay, Michelle Neudeck, and Silvia E.
Newell**

Sandusky Results: Potential Ammonium Uptake and Regeneration

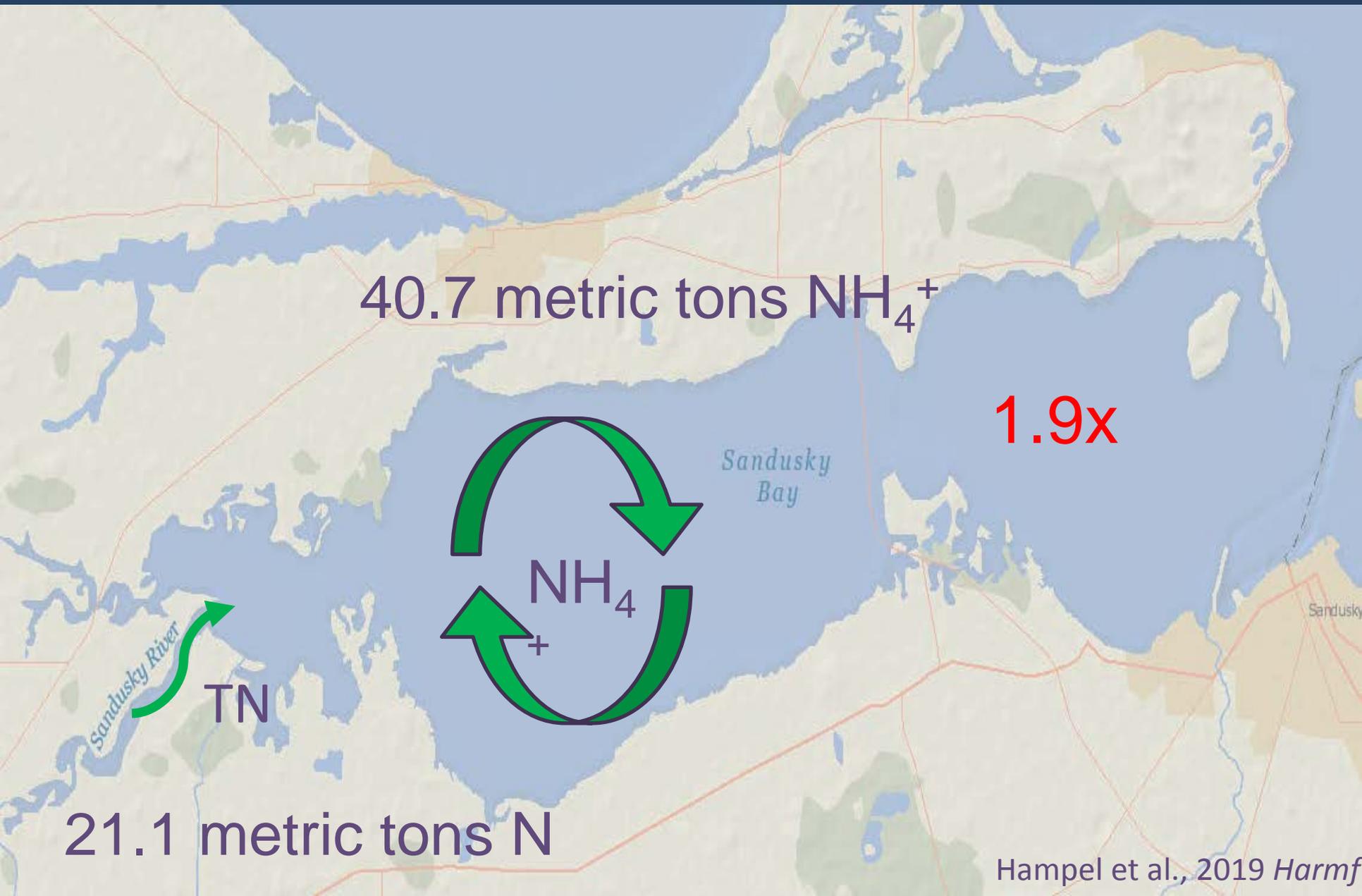
NH_4^+ uptake in light



NH_4^+ Regeneration

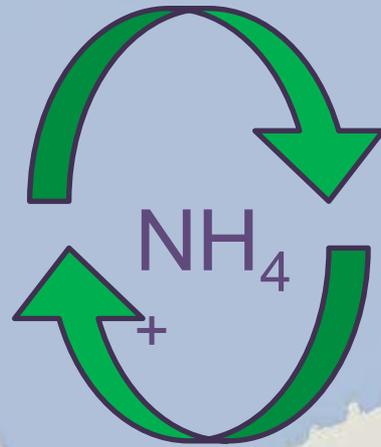


June 5th, 2017



40.7 metric tons NH_4^+

1.9x



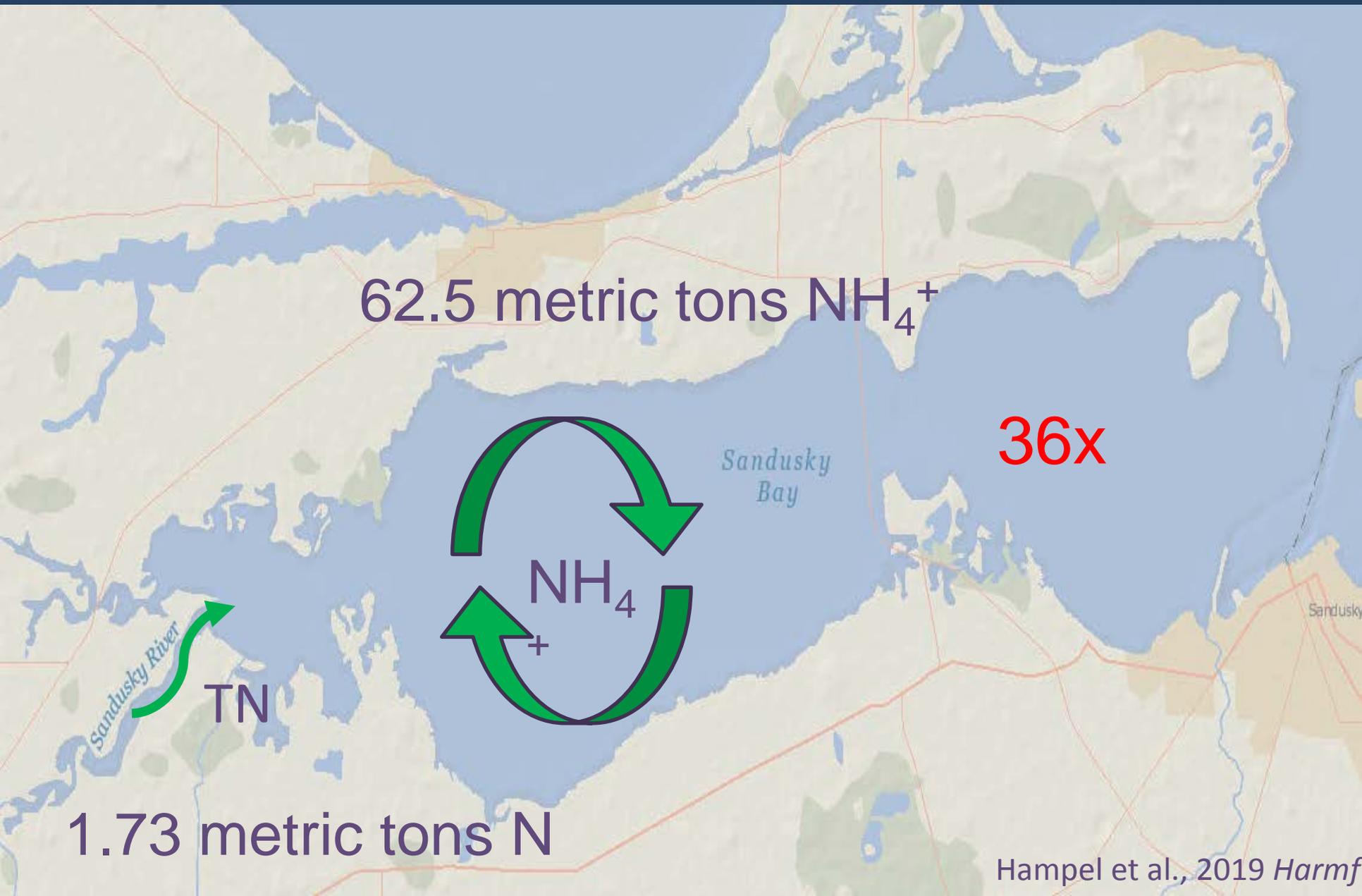
Sandusky Bay

Sandusky River

TN

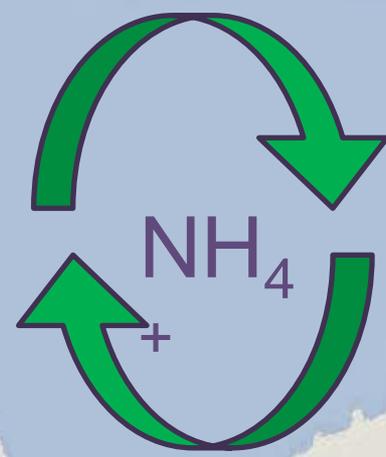
21.1 metric tons N

July 31st, 2017



62.5 metric tons NH₄⁺

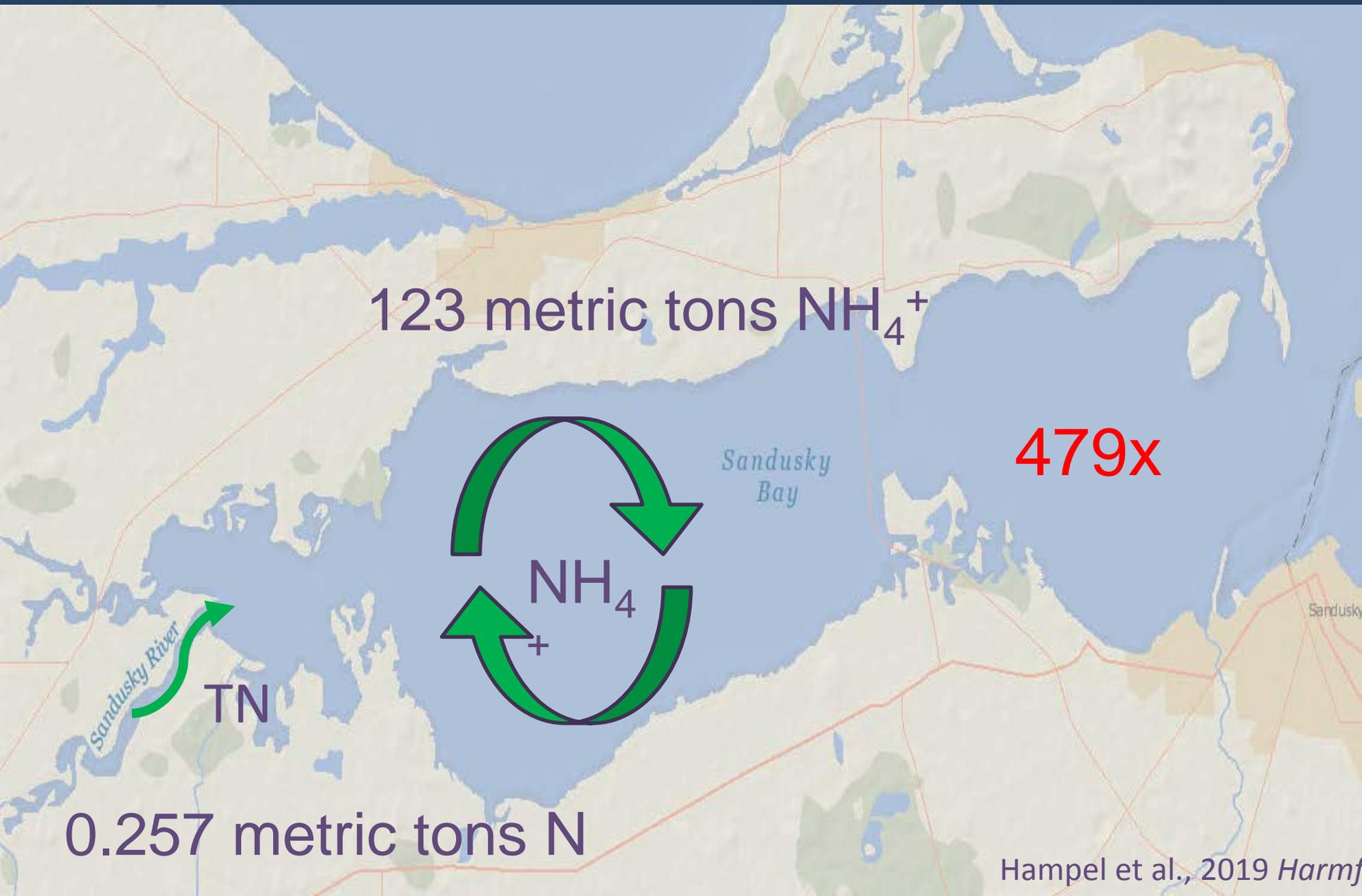
36x



TN

1.73 metric tons N

August 14th, 2017

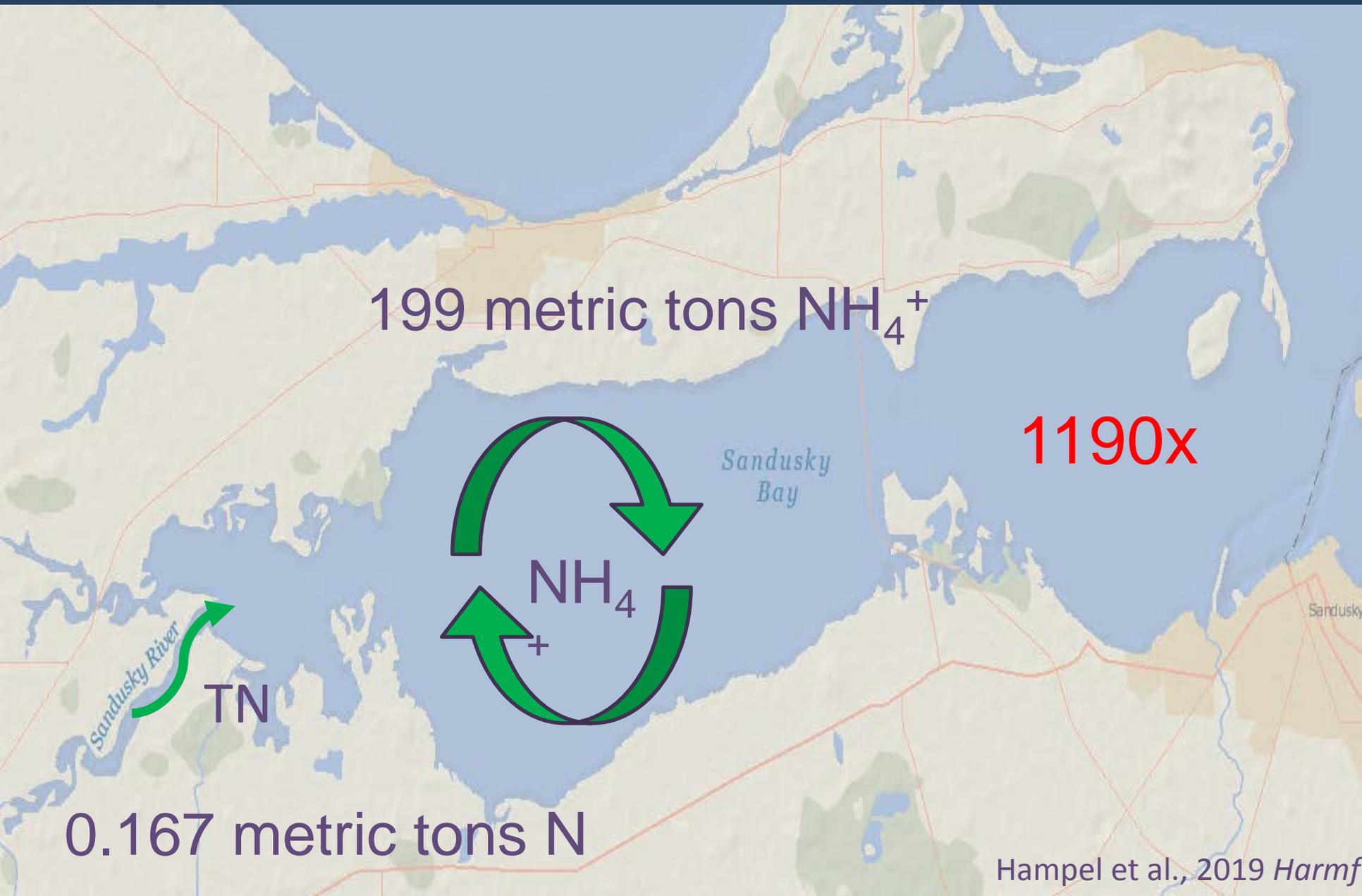


0.257 metric tons N

123 metric tons NH₄⁺

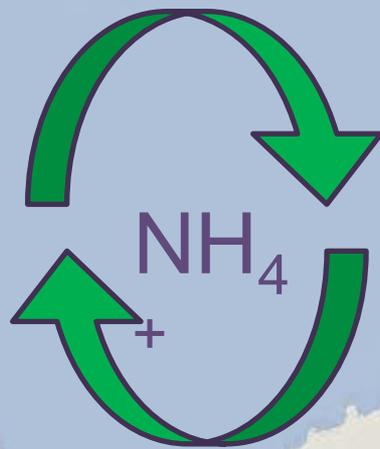
479x

August 28th, 2017



199 metric tons NH_4^+

1190x



TN

0.167 metric tons N



Sandusky Bay: Ammonium Regeneration

- Regeneration in the bay increases throughout the summer suggesting that toward the end of summer the bloom relies heavily on regenerated NH_4^+ .
 - With undetectable DIN in the water column, regeneration sustains the bloom
- When extrapolated to the whole bay volume, daily NH_4^+ regeneration exceeded daily TN loadings at all sampling events.
 - Useful tool for N management practices and nutrient regulations.

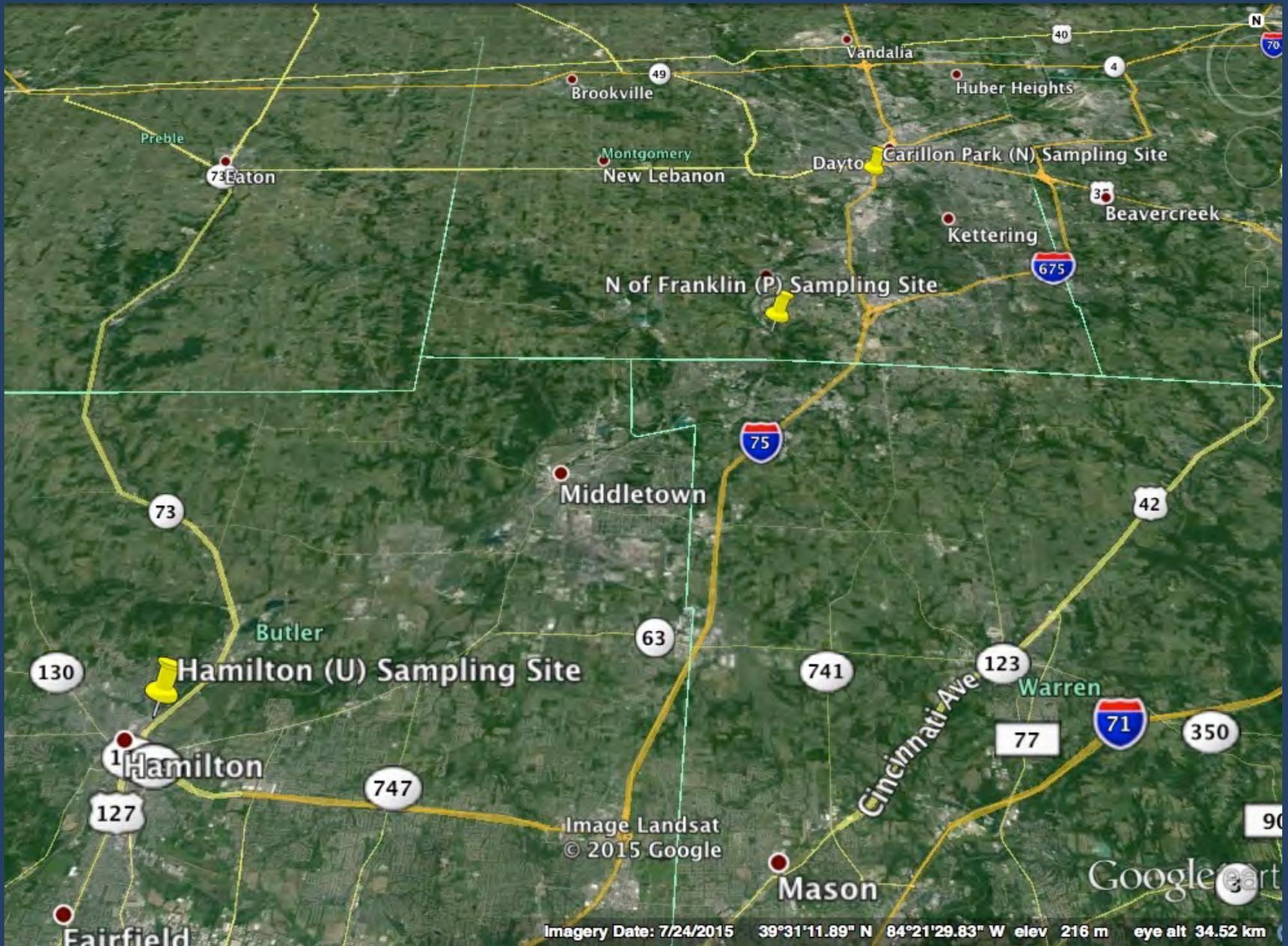


Eutrophication in the Great Miami River: To What Extent Can Sediment Nitrogen Loss Compensate for Nutrient Over-enrichment?

Lee Slone, M.Sc. student – WSU

Slone et al. 2018

Slone et al., 2018

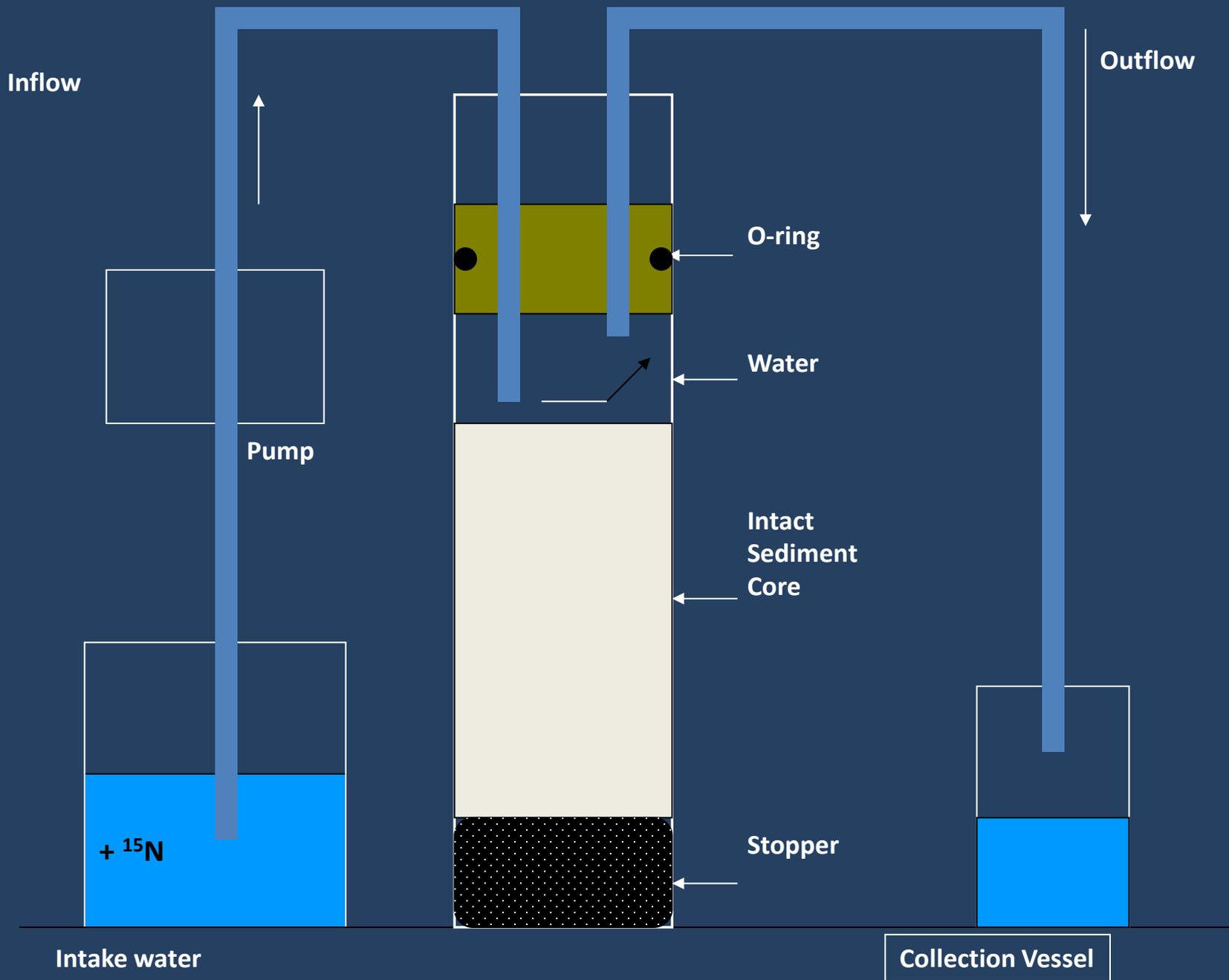


Collect intact sediment cores and near-bottom water for continuous-flow incubations to measure SWI N fluxes and transformations.



Photos courtesy of Nate Christopher at Fondriest Env.







Denitrification



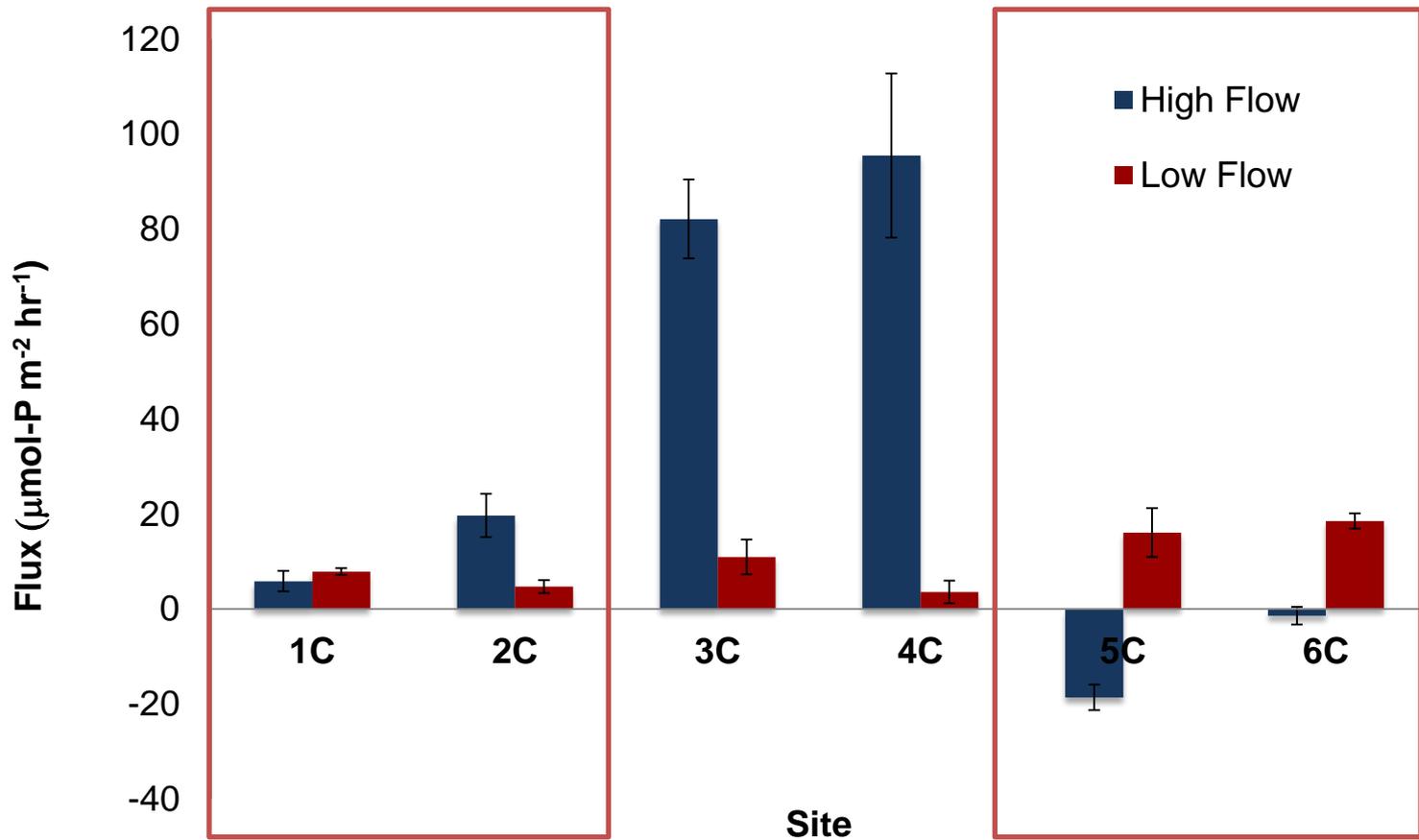
- Requires anoxia
- Pathway ends at N_2O more often under low oxygen
- Removes excess N from ecosystem
- Critical ecosystem service in eutrophic systems!

Dissolved gas fluxes

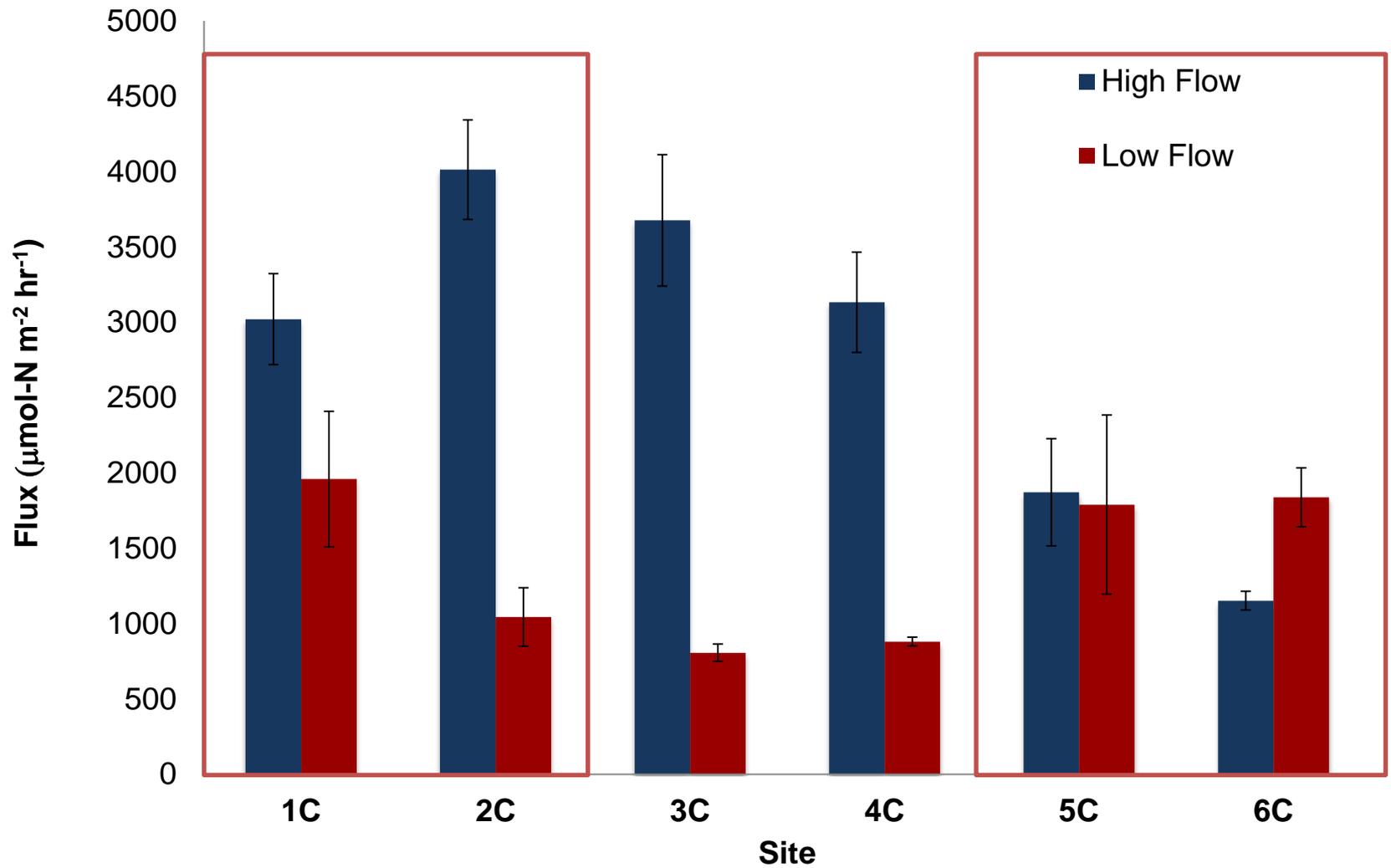
- Dissolved gases ($^{28}\text{N}_2$, $^{29}\text{N}_2$, $^{30}\text{N}_2$, O_2 , Ar)
- MIMS (Kana et al. 1994, An et al. 2001)



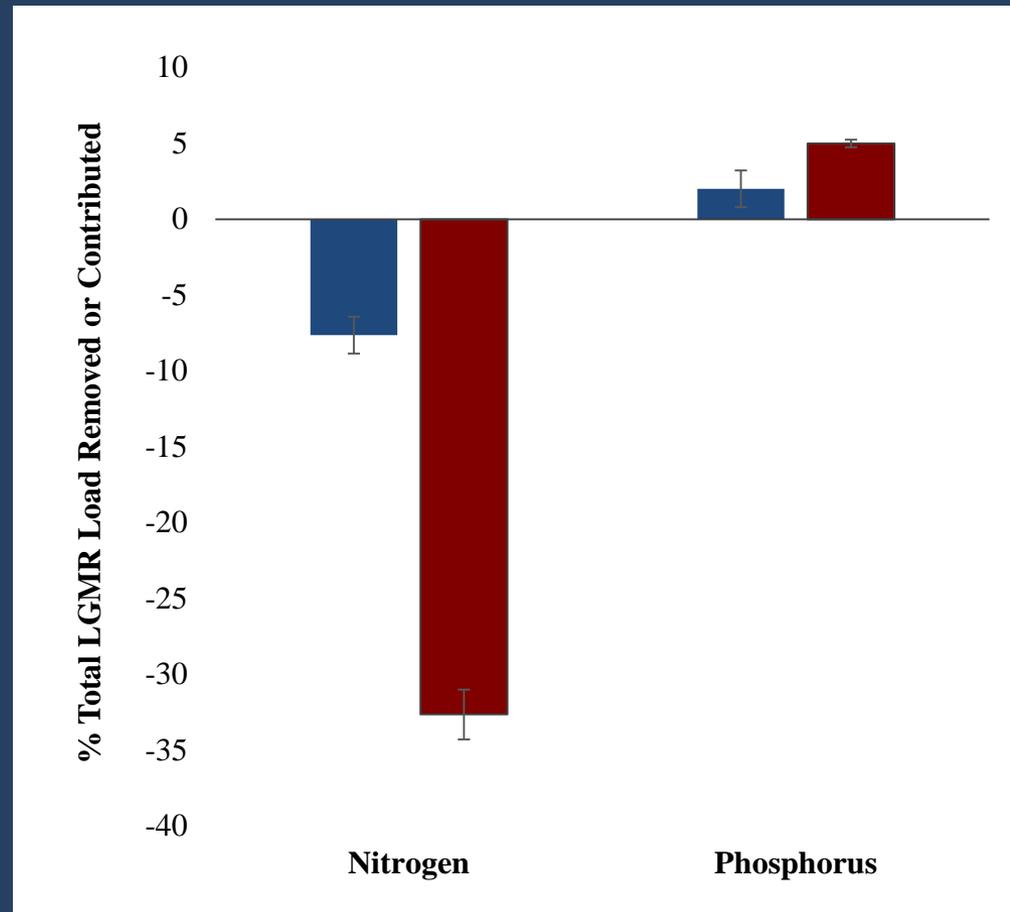
Nutrient flux: PO₄



Excess $^{28}\text{N}_2$ flux (Denitrification)



Impact on River Nutrient Load

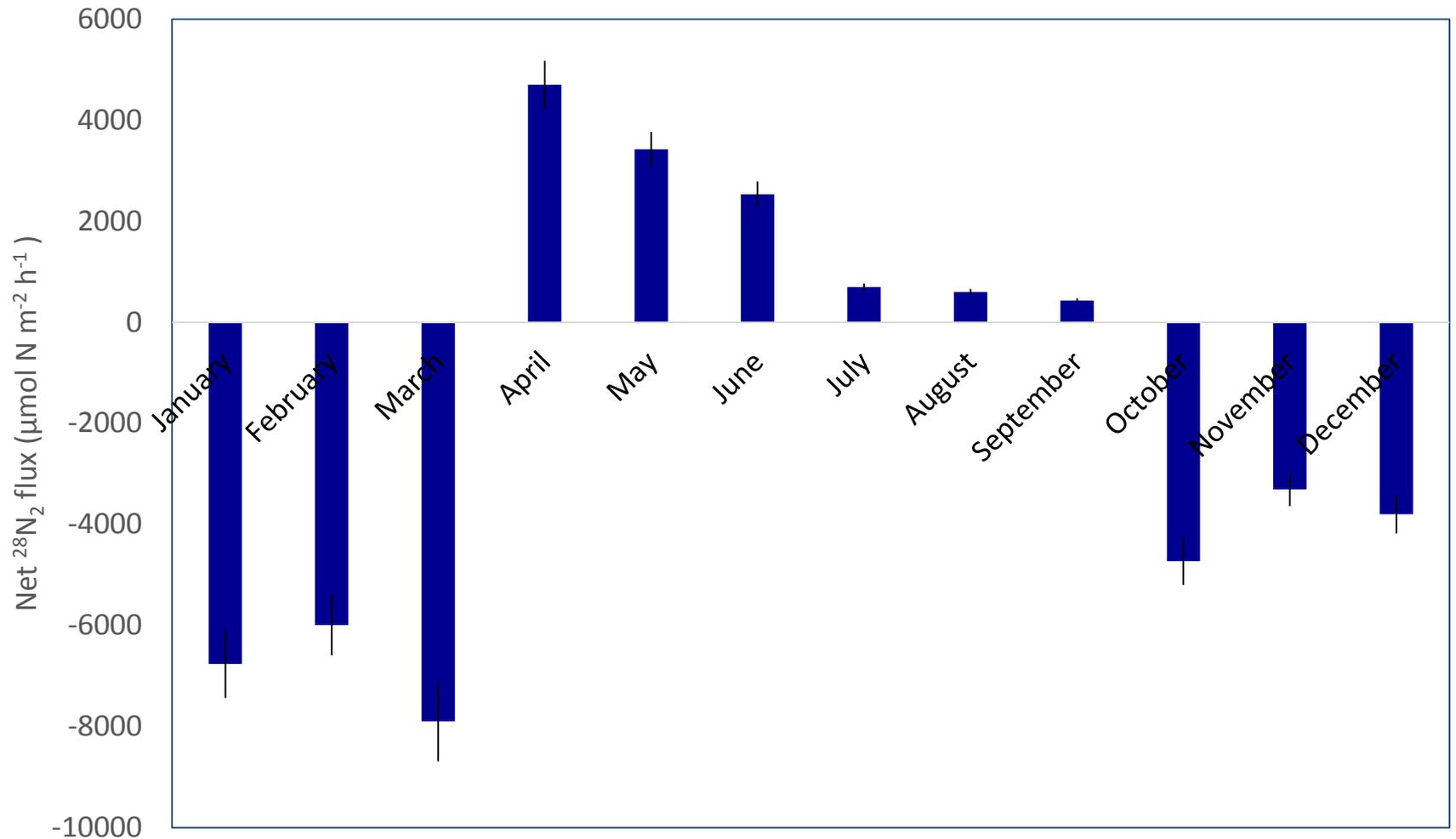


Slone et al., 2018

LGMR Conclusions

- Denitrification removed nitrate at very high rates during high river flow, but N removal was still exceeded by external inputs (to a lesser extent during low flow).
- Sediments were a source of bioavailable ammonium, potentially contributing to local algal blooms, but were a strong nitrate sink and overall net N sink.
- River sediments were a P source, contributing an additional 2 - 6 % to the external P load from the agricultural and urban watershed.
- These results support calls for a dual nutrient (N & P) management approach to control eutrophication in inland waters and coastal marine systems.

Net Denitrification or Nitrogen Fixation Lower Great Miami River 2017-2018



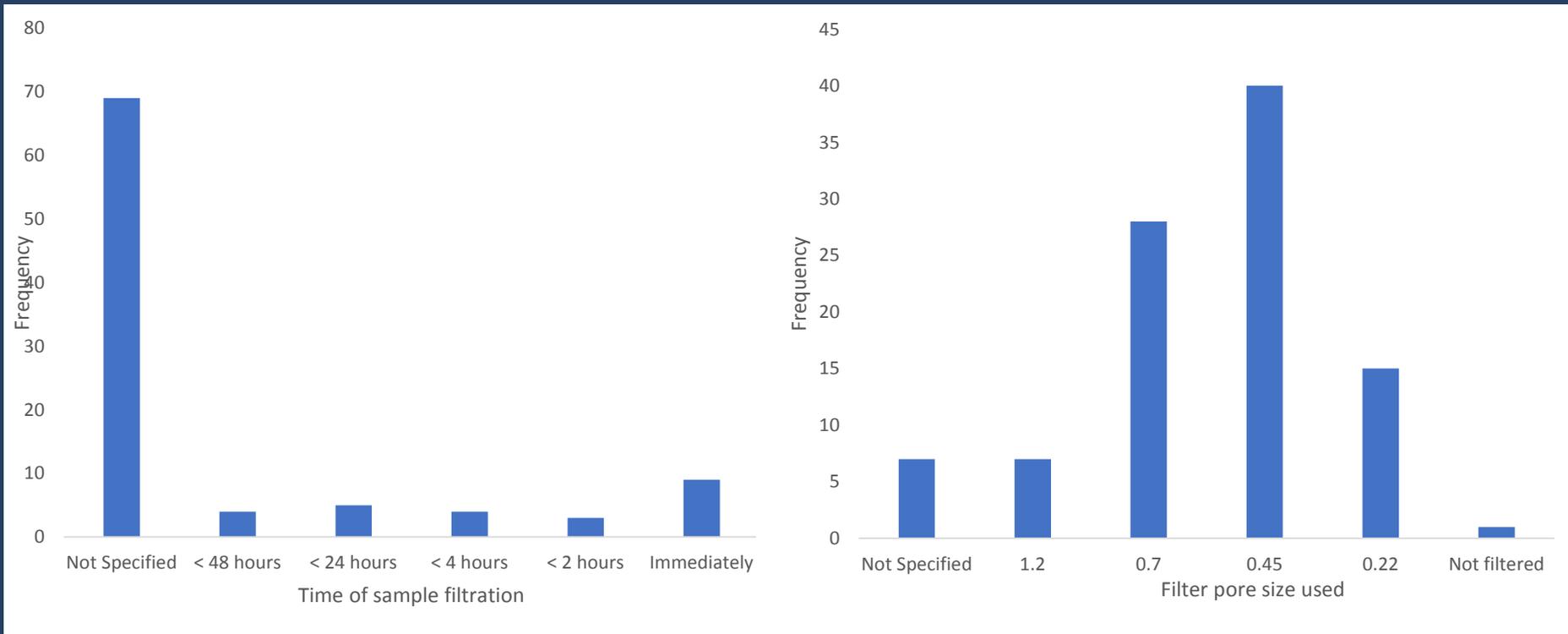
Slone et al. 2018

Slone et al., 2018

Nutrient sample collection

- Megan Reed et al.

Journal article survey of nutrient sampling methods



Why does accurate nutrient data matter?

- Using data from Hampel et al 2018:

Uptake: $0.759 \mu\text{mol L}^{-1} \text{h}^{-1}$

Regeneration: $0.337 \mu\text{mol L}^{-1} \text{h}^{-1}$

Ambient concentration of ammonium (NH_4^+): $0.33 \mu\text{M}$

An unfiltered water sample stored in the dark would have:

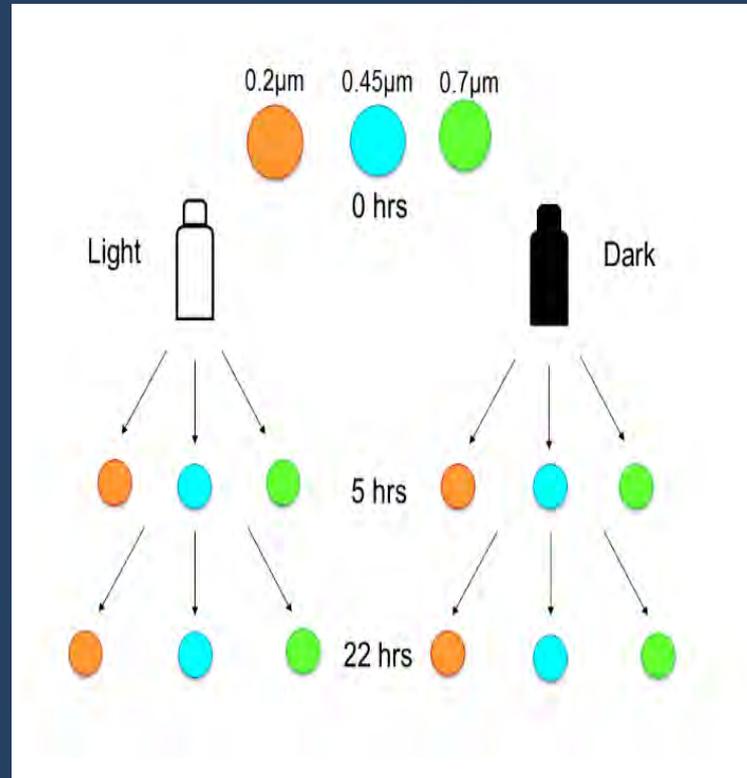
$[\text{NH}_4^+]$: $0.119 \mu\text{M}$ after 30 minutes

$[\text{NH}_4^+]$: $0 \mu\text{M}$ after only 47 minutes!

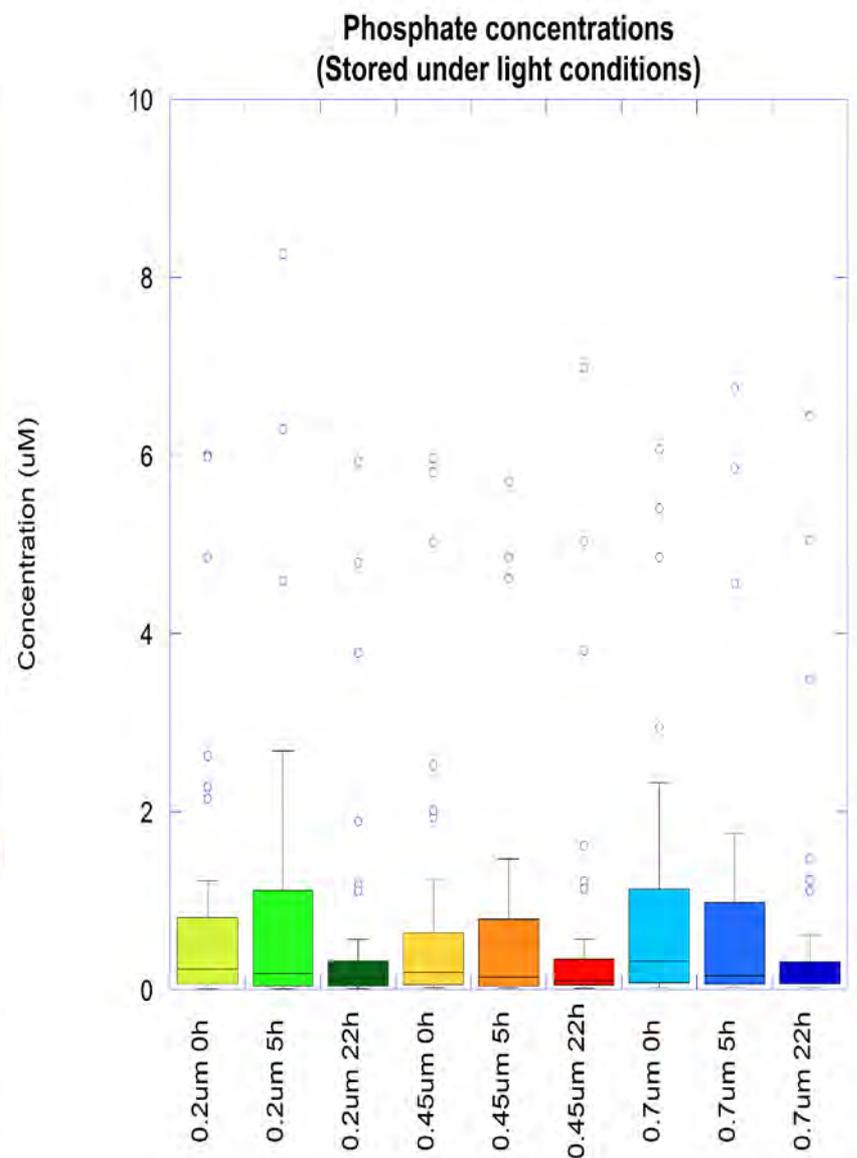
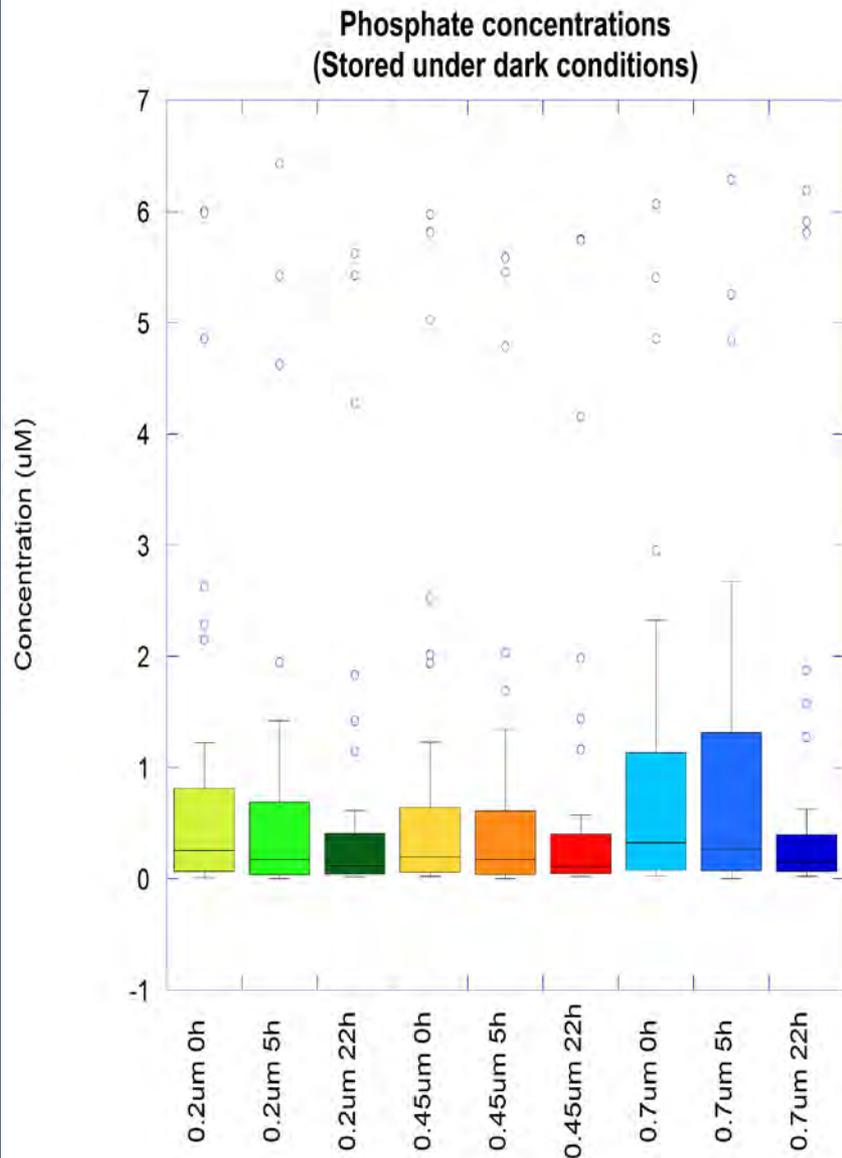
Methods- Study site



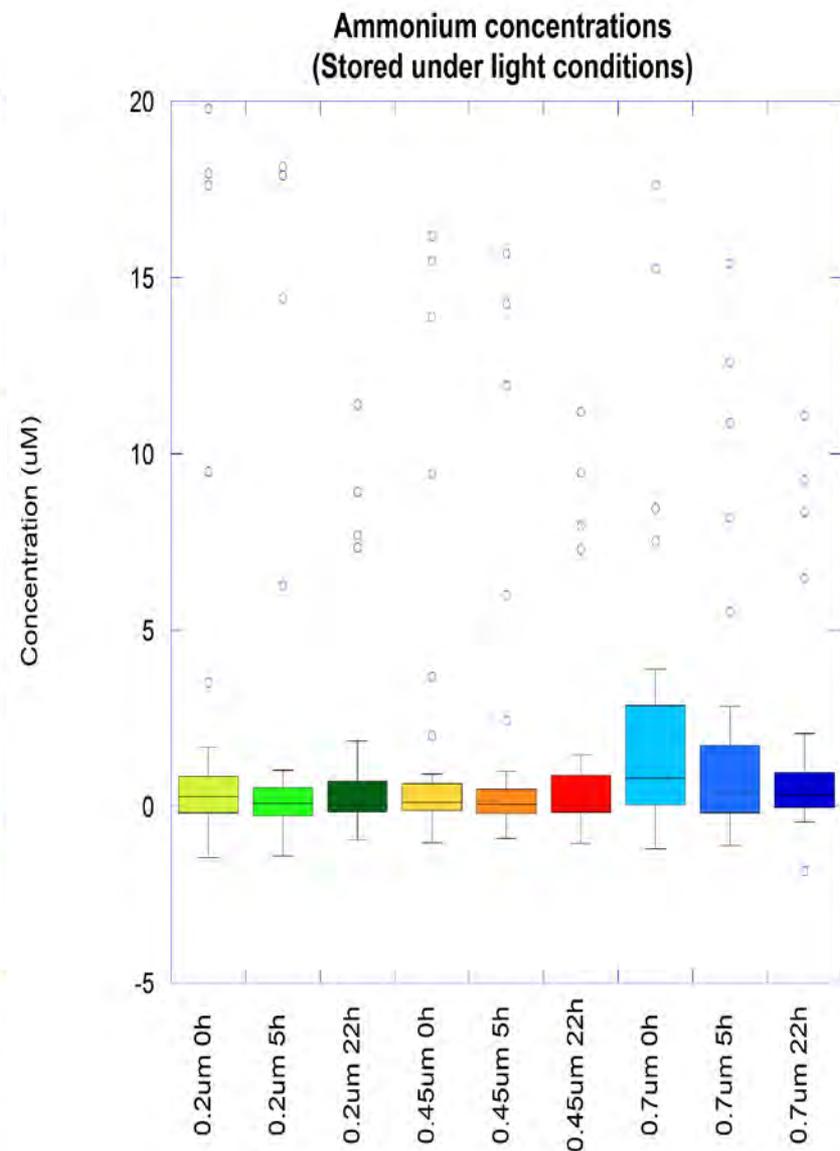
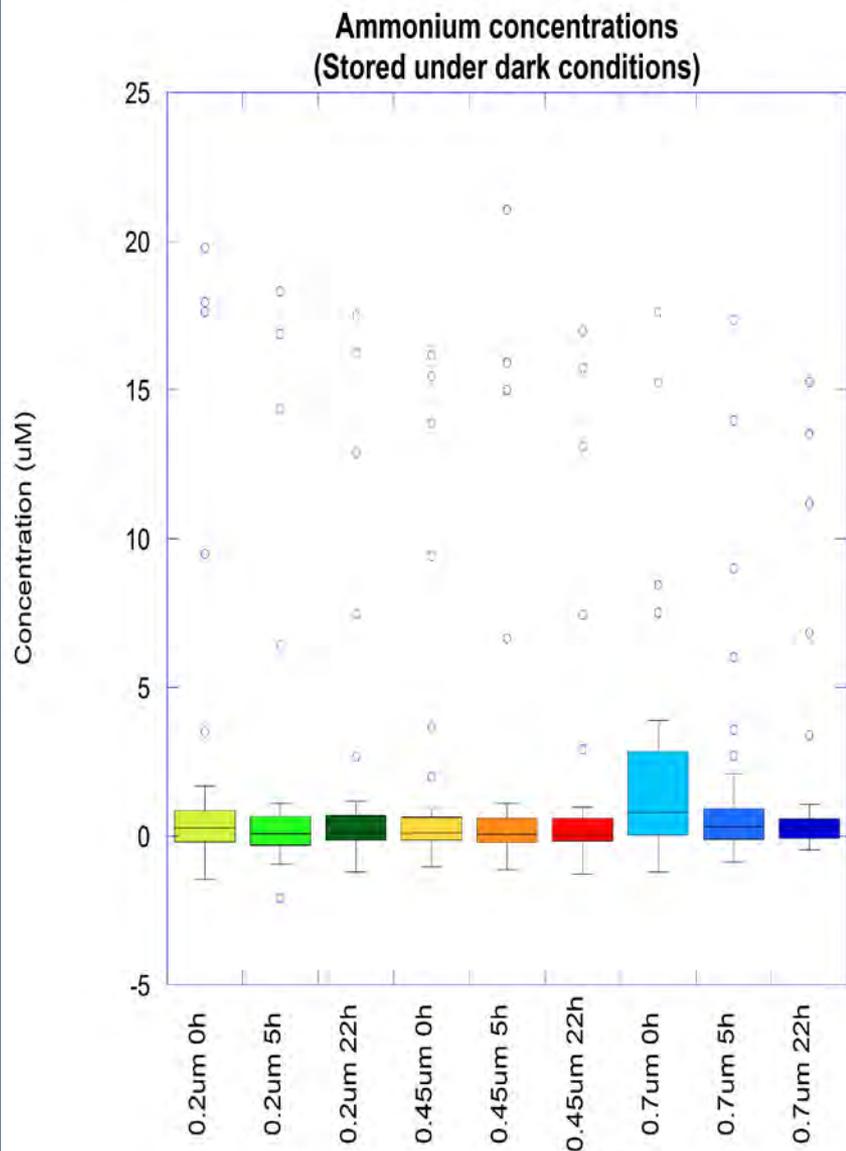
Sample collection



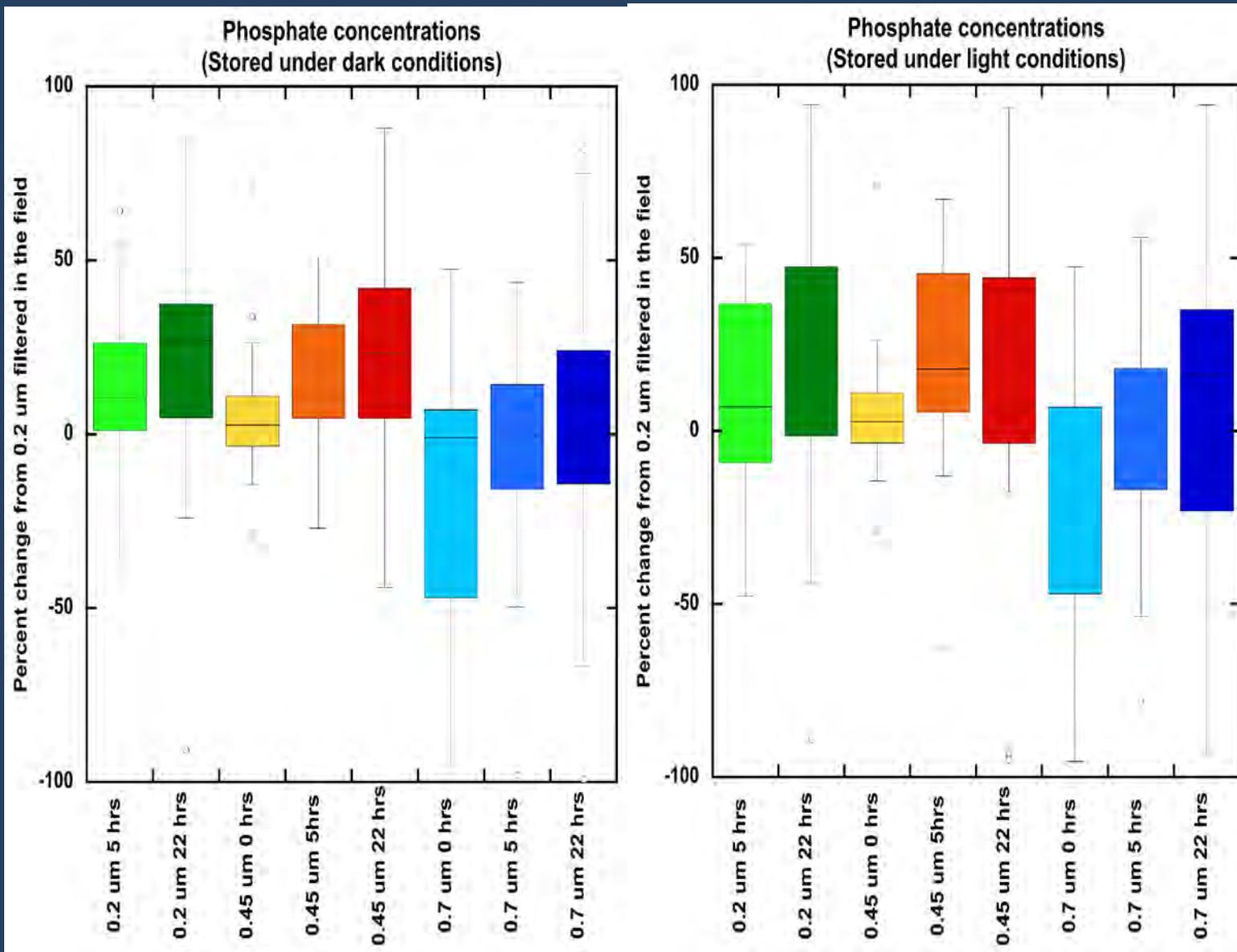
Results- Phosphate concentrations



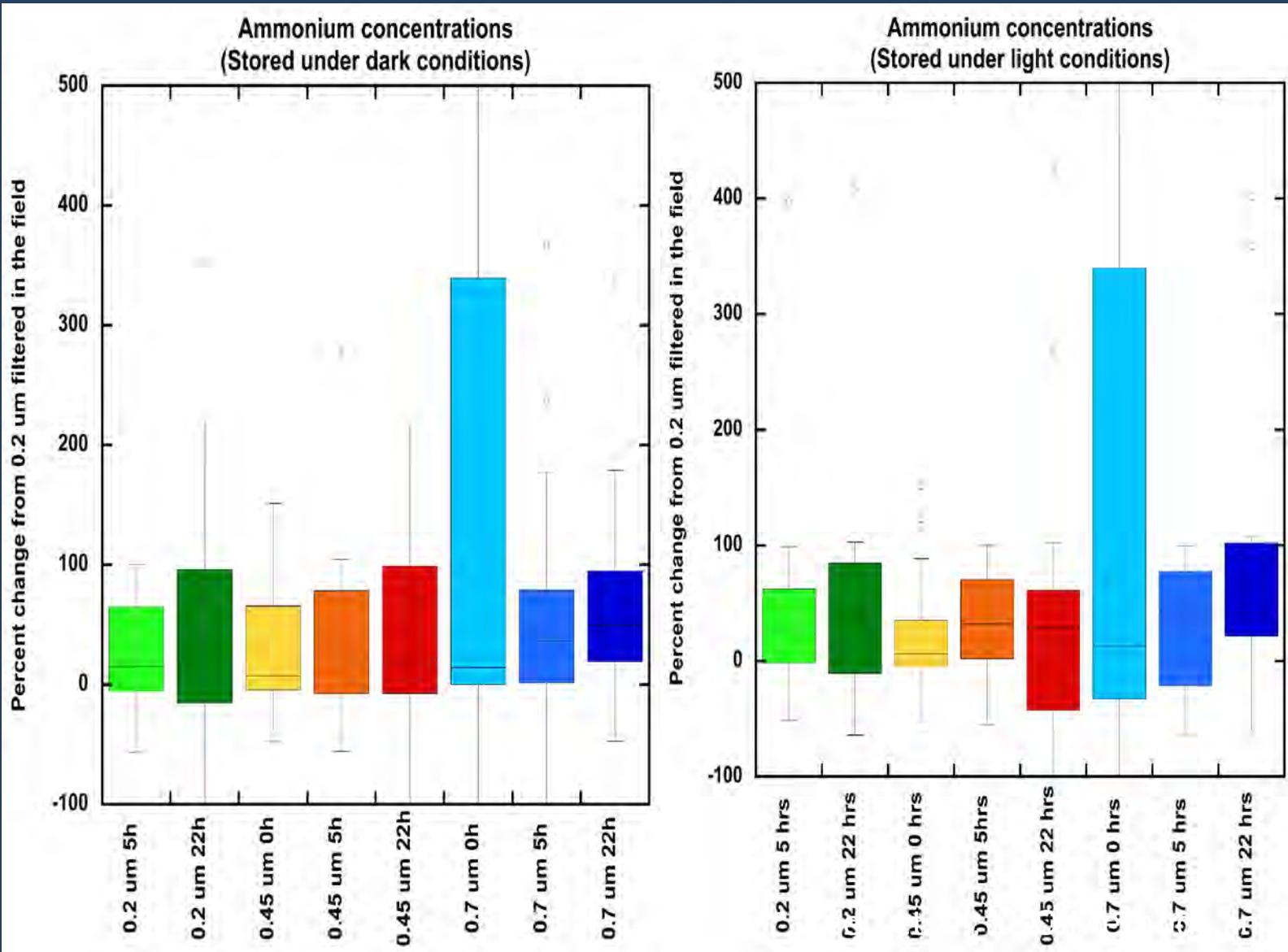
Ammonium concentrations



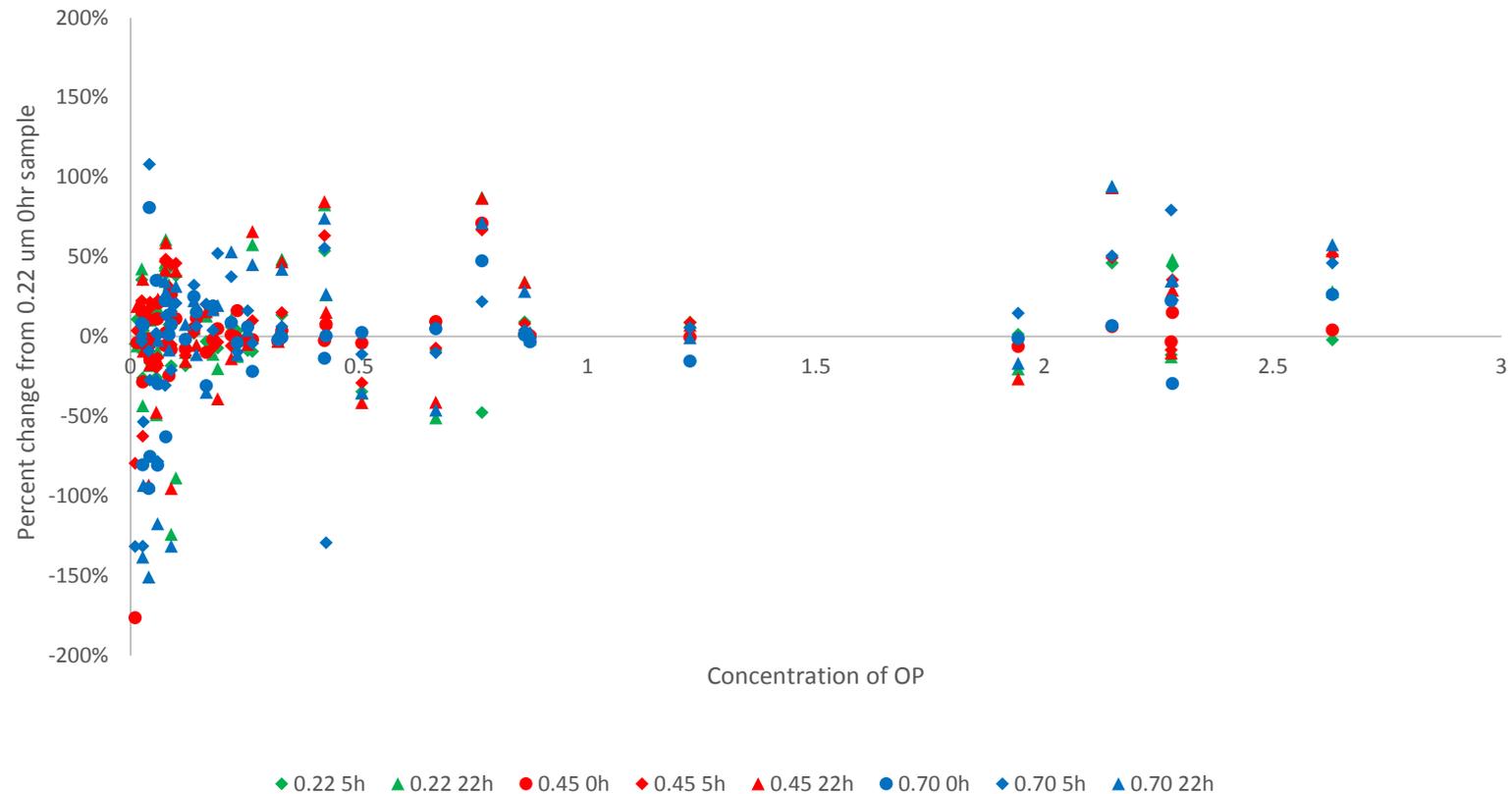
Percent change of phosphate concentrations



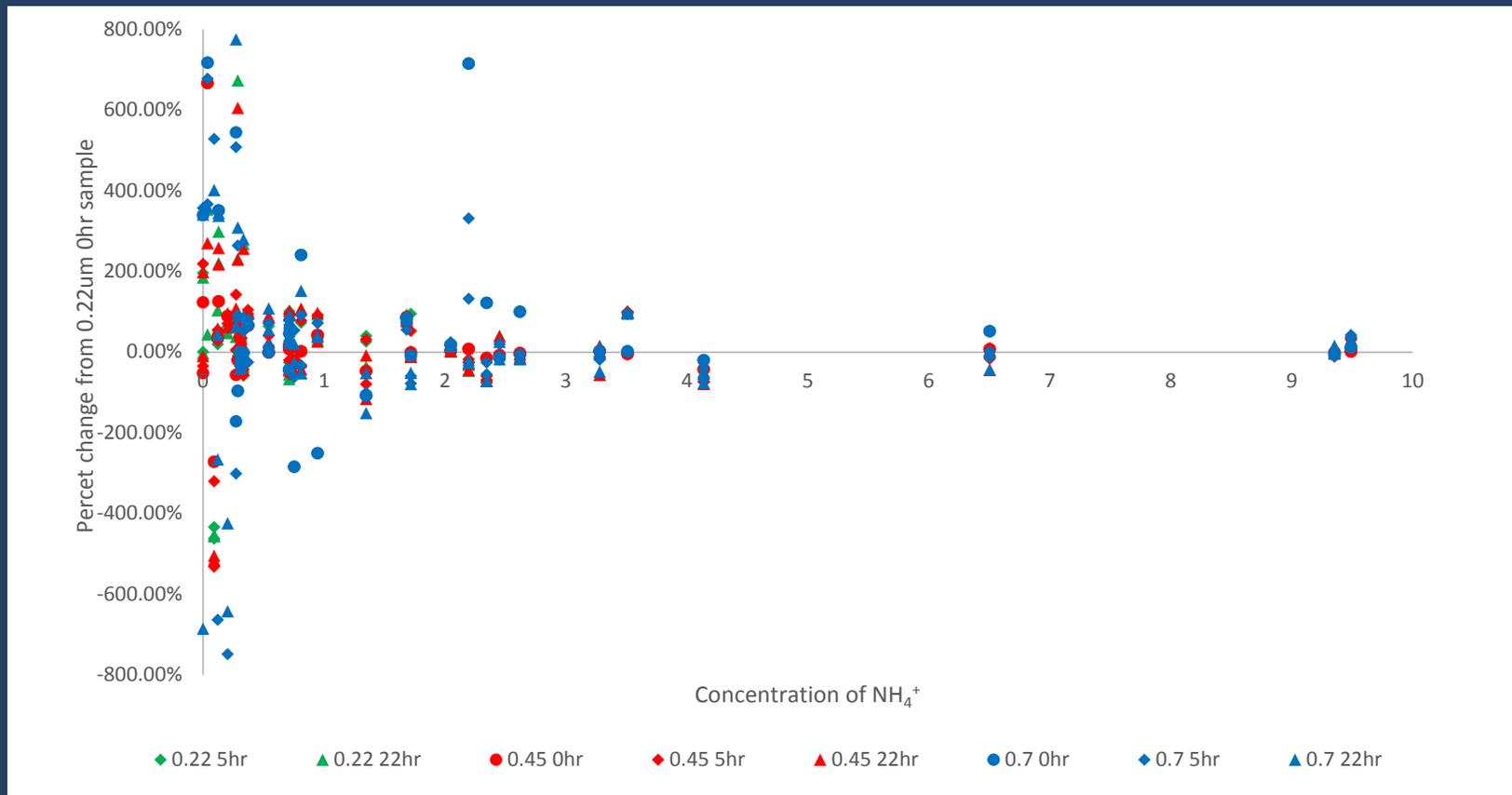
Percent change of ammonium concentrations



Concentration of P against percent change from 0.22 μM sample filtered in the field



Concentration of N against percent change from 0.22 μM sample filtered in the field



Sample Collection Conclusions

- Failing to filter samples in the field can lead to significant changes in the concentration compared to the ambient value
- Filtering in the field with 0.70 μm filters has the most significant impact on nutrient concentration
- More specific filtering methods should be considered in order to ensure the accuracy of reporting nutrient monitoring data

Thank you!

- Zhang Lu
- Xu Hai
- Zhu Guangwei
- Yang Longyuan
- Zhu Mengyuan
- Gao Guang
- Tang Xiangming
- Hans Paerl
- Steve Wilhelm
- Liu Kaijun
- Justin Chaffin/
OSU Stone Lab
- Hammerschmidt
Lab
- Tim Davis/ NOAA
GLERL
- Sue Watson/Env.
Canada
- Miami
Conservancy
District

